Exxon Valdez Oil Spill Restoration Project Final Report

Injury to Salmon Eggs and Preemergent Fry in Prince William Sound

> Restoration Project 93003 Final Report

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Study History: This study originated in March of 1989 and continued through February of 1991 as Natural Resource Damage Assessment Fish/Shellfish Study 2. The project consisted of fall embryo sampling and spring preemergent fry sampling in oil contaminated and unimpacted streams to determine if the *Exxon Valdez* oil spill had an impact on incubating pink salmon. This work continued in 1992 as Restoration Study R60C. At that time the project was expanded to include the previously described field sampling as well as 1) laboratory evaluation of field results through the controlled incubation of pink salmon embryos on oiled substrate conducted by the National Marine Fisheries Service **Auke** Bay Lab, 2) **an** experiment designed to evaluate whether the results observed in the field were due to characteristics of the spawning population or stream characteristics by the Alaska Department of Fish and Game, and 3) a search for evidence of genetic damage by the Alaska Department of Fish and Game's Genetics Laboratory. This project, 93003, is a continuation of R60C. Final reports have been written for Fish/Shellfish Study 2 and Restoration Study R60C.

<u>Abstract</u>: We examined the possibility that the differences in pink salmon embryo mortality observed in the field in past years was due to traits carried by the parents. Gametes were collected from adults in spawning condition as they amassed on or near the spawning ground from eight oil contaminated and eight unimpacted streams. The resulting pink salmon embryos from oil contaminated streams showed elevated mortalities when compared to the embryos from unimpacted streams. No difference in embryo to preemergent fry survival for pink salmon incubating during the winter of 1992-1993 was detected between oil contaminated and unimpacted streams.

Key Words: Onchorhynchus gorbuscha, embryo mortality, overwinter survival, eggs, preemergent fry, pink salmon, Prince William Sound.

<u>Citation</u>:

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EXECUTIVE SUMMARY

This study is a continuing project designed to monitor recovery of pink salmon *Oncorhynchus gorbuscha* populations in Prince William Sound which were impacted by the *Exxon Valdez* oil spill. Embryo mortality and embryo to preemergent fry survival have been examined in intertidal and upstream areas of oil contaminated and unaffected (control) streams since the spring of 1989. This report covers work performed between March 1, 1993 and September 30, 1993.

Embryo mortality was significantly greater in oiled streams in 1989 and 1990 with the differences observed in all intertidal areas in 1989 and in the highest intertidal area in 1990. These results were consistent with observations on intertidal oiling from other studies: among oiled streams, all intertidal areas were contaminated in 1989 whereas in 1990 oil remained only in the upper intertidal zone.

The 1991 evaluation demonstrated a very significant difference in embryo mortality between oil contaminated and control streams with the differences in both the intertidal and upstream zones. The pink salmon which spawned during the fall of 1991 were from the 1989 brood year, the brood year which incubated in oiled gravels during the fall of 1989 and spring of 1990. This finding was unexpected and raised questions as to the possibility of an oil induced damage manifesting itself in the form of functional sterility and that these damages may be transmitted genetically. A pattern of embryo mortality similar to, but not as extreme as 1991 was observed in 1992.

We examined the possibility that the differences in pink salmon embryo mortality observed in recent years was due to a trait carried by the parents. Gametes were collected from adults in spawning condition as they amassed on or near the spawning grounds from eight oil contaminated and eight control streams. The gametes were flown to the Armin F. Koeming hatchery in southwest Prince William Sound where intrastream crosses were made and the resulting embryos from each stream placed in a common incubator. The resulting pink salmon embryos from oil contaminated streams showed elevated mortalities when compared to the embryos from control streams.

No difference in embryo to preemergent fry survival has been detected. We expected survival to be reduced in oil contaminated streams for all years examined given that an increase in embryo mortality was already detected; this has not been the case. We suspect that unexpected changes in stream characteristics prevented us from sampling the same areas or populations for embryos in the fall and fry in the spring. It is also common for intertidal stream segments to migrate along the beach especially if the beach is exposed to winter storms. The magnitude of these changes was unexpected when this study was designed and initiated.

INTRODUCTION

Wild salmon play a major role in the Prince William Sound ecosystem while also contributing to the region's commercial fisheries. Migrating salmon *fry* are an important food source in the spring for various mammals, birds, and fishes. Marine mammals prey on the ocean life stages of Pacific salmon while terrestrial mammals and birds, such as bears, river otters, eagles, and gulls depend on salmon for a large portion of their summer diet. Salmon also provide a pathway for transferring nutrients from marine ecosystems to near-shore and terrestrial ecosystems. In recent years, commercial catches of wild salmon have ranged from 10 to 15 million pink salmon and from 0.8 to 1.5 million chum salmon.

Up to 75% of spawning pink *Oncorhynchus gorbuschu* and chum salmon O. *keta* in Prince William Sound use intertidal areas (Helle et al. 1964). These areas are highly susceptible to contamination from marine oil spills. Moles et al. (1987) and Rice et al. (1975) found that pink salmon embryos and preemergent fry were adversely affected by exposure to crude oil and that the affect was most acute in intertidal environments. The March 24, 1989 spill from the *Exxon Valdez* occurred just prior to the spring migration of salmon fry and contaminated many intertidal spawning areas in central and southwest Prince William Sound.

Embryo mortality was significantly greater in oiled streams in 1989 and 1990 (P=0.004 and P=0.023) with the differences being in all intertidal areas in 1989 and in the highest intertidal area in 1990 (Figure 1; Sharr et al. 1994a). These results were consistent with the observations of Wolfe et al. (*In press*) on intertidal oil contamination: among oiled streams, all intertidal areas were contaminated in 1989 whereas in 1990 oil remained only in the upper intertidal zone.

The 1991 evaluation demonstrated a very significant difference in embryo mortality (P=0.003) between oil contaminated and control streams (Figure 1; Sharr et al. 1994a). The pink salmon which spawned during the fall of 1991 were from the 1989 brood year, the brood year which incubated in oiled gravels during the fall of 1989 and spring of 1990. This finding was unexpected and raised questions as to the possibility of an oil induced damage manifesting itself in the form of functional sterility and that these damages were transmitted genetically. A pattern of embryo mortality similar to but not as extreme as 1991 was observed in 1992 (P=0.010; Figure 1; Sharr et al. 1994b). We felt the 1992 results supported the genetic damage hypothesis since fewer fish were most likely impacted in 1990; consequently, the subsequent difference observed in 1992 would not be as great.

These field findings were interesting, but not definitive proof that oil contamination was responsible for the increased embryo mortality or that there was a link in damages between generations. It was possible that the observed discrepancies were due to environmental differences between streams which we were unable to account for due to the observational nature of the study.



Figure 1. Pink salmon embryo mortality observed during past field sampling. Solid circles indicate the means for oil contaminated streams (n=10) while open circles identify control streams (n=15).

No difference in embryo to preemergent fry survival between oil contaminated and control streams has been observed since the initiation of the study in 1989 (Figure 2; Sharr et al. 1994a and 1994b). We expected embryo to preemergent fry survival to be reduced in oiled streams given an increase in embryo mortality was already detected. This result can potentially be explained by (1) compensation in the intergravel environment, or (2) problems in the experimental design. Geiger et al. (*Inpress*) found no evidence to suggest that compensation in the intergravel life stages is playing a role in determining the number of emerging *fry* for the years in the study. We believe that the experimental design is inadequate for detecting differences in embryo to preemergent fry survival.

We have continued to collect preemergent fry data. The Alaska Department of Fish and Game has collected preemergent fry density data from 25 Prince William Sound streams since 1961. This data has been primarily used to forecast future returns of wild pink salmon. The data collection for these 25 streams is funded by the State of Alaska general operating budget. The preemergent fry portion of this project collects data from an additional **23** streams in the oil contaminated area of the sound. The additional data is used to monitor oil damaged streams as well as provide a database for future wild pink salmon forecasts.

This study was initially designed to monitor the effect of intertidal oiling on pink salmon embryo mortality and embryo to preemergent fry survival. The project was amended during the summer of 1992 to evaluate the genetic damage hypothesis. Two controlled experiments were designed and initiated; 1) an experiment to evaluate whether the results observed in the field are due to physical stream conditions such as stream orientation, drainage characteristics, or weather differences (administered by Alaska Department of Fish and Game); and 2) a controlled oiling experiment designed to verify the field findings and evaluate the genetic damage hypothesis (administered by National Marine Fisheries Service).



Figure 2. Pink salmon embryo to preemergent fry survival observed during past field sampling. Solid circles indicate the means for oil contaminated streams (n=10) while open circles identify control streams (n=15).

OBJECTIVES

Recovery Monitoring of Injury to Pink Salmon Eggs and Preemergent Fry in Prince William Sound

- 1. Estimate the density by tide zone of preemergent fry in **48** streams and embryos in 31 streams using numbers of live and dead embryos and fry.
- 2. Estimate embryo mortality and overwinter survival of pink salmon embryos in both oiled and unoiled streams.
- 3. Assess any loss in adult production from changes in overwinter survival using the results of **NRDA** F/S Studies 1, 2, 3, and 4.
- **4.** Identify potential alternative methods and strategies for restoration of lost use, populations, or habitat where injury is identified.

Verification of Injury to Pink Salmon Gametes in Prince William Sound

1. Determine whether the increased pink salmon embryo mortalities observed in oiled streams by Sharr et al. (1994a and 1994b) can be attributed to the physical characteristics of the study streams.

METHODS

Recovery Monitoring of Injury to Pink Salmon Eggs and Preemergent Fry in Prince William Sound

Study Sites

This project concentrated on the southwestern portions of Prince William Sound; although, streams from Montague Island and eastern Prince William Sound were sampled to provide a sound wide perspective (Figure 3). Forty-eight streams were sampled between March 12 and May 13, 1993, for preemergent salmon fry. Twenty-five of these have been historically sampled to provide data for forecasting future adult pink salmon returns. These streams were



Figure 3. Streams examined by the 1989, 1990, 1991, 1992, and 1993 pink salmon preemergent fry surveys.

selected for the following reasons:

- 1. They contribute a large proportion of the wild return of pink and chum salmon to Prince William Sound.
- 2. They have significant spawning populations in both odd and even years.
- 3. They **are** representative of the spatial distribution of spawning escapement in Prince William Sound.
- 4. They **are** accessible for sampling in most years.

The additional 23 streams have been sampled since the 1989 *Exxon Valdez* oil spill. These additional streams were primarily located in the southwestern Sound in the oil impacted **area.**

Sample Design

The methods used for both embryo and preemergent fry sampling were similar to those described by Pirtle and McCurdy (1977) and Sharr et al. (1994a and 1994b). Sampling was stratified by tide zone to control for possible differences in embryo mortality or overwinter survival due to differences in salinity, temperature, predation, oiling, or a combination of these factors. Zone boundaries were established with a surveyor's level and stadia rod and staked prior to fry sampling. Four zones, three intertidal and one above tidal inundation, were sampled whenever possible, for each stream: 1.8 - 2.4 m, 2.4 - 3.0 m, 3.0 - 3.7 m above mean low water, and upstream of mean high tide (3.7 m). No sampling was done below the 1.8 - 2.4 m zone because survival was expected to be low (Helle et al. 1964). Upstream sample transects were often within the reach of extreme high tides (3.7 - 4.6 m) since spring ice and snow conditions often limited the extent of upstream sampling for preemergent fry.

Separate linear transects were established for each zone on the embryo and preemergent fry surveys. Although most transects were 30.5 m long, some were shorter due to steep stream gradients. Transects were placed in riffle areas where spawning was observed during escapement surveys conducted for NRDA F/S Study 1. Transects ran diagonally across the river: Fry survey transects started downstream against the right bank and moved upstream to the left bank while embryo survey transects started downstream against the left bank and moved upstream to the right bank. This placement of embryo and fry transects reduces sampling overlap and the influence of fall embryo sampling on spring fry abundance. A map drawn for each stream indicated the tide zones and transect locations in relation to major landmarks. Each embryo transect was photographed and marked with surveyor's flagging to assure that fry transects could be located in the Same area of the stream. This was done to better estimate embryo to fry survival within each sample zone.

Fourteen circular digs, each 0.186 m^2 , were systematically made along each transect. The number of digs was a compromise between reducing variance and the practicality of conducting the study. Fewer digs were completed on narrow stream channels to avoid excessive sampling of the stream. Streams that split into two or more channels within a zone

were sampled either by allocating digs among channels based on spawner distribution observed during NRDA F/S Study 1 or, where spawner distribution was unknown, by an equal allocation.

The following data were collected for each tide zone transect during both embryo and fry sampling:

- 1. Sample date.
- 2. Sample tide zone.
- **3. Start** and stop time for the tide zone transect.
- **4.** Numbers and condition (live or dead) of fry and embryos by species for each dig.
- 5. A subjective estimate of the overall percent yolk sac absorption for fry in each dig.

Pink salmon fry were differentiated from chum salmon fry by their smaller size and lack of parr **marks.** An embryo was considered dead if it was opaque or discolored with concentrations of lipids. Sampling often **killed** fry (especially newly hatched fry), so fry were only considered dead if decomposition was evident.

Data Analysis

Densities of live preemergent fry (\hat{F}_{ij}) for stream i, zonej in m² were estimated by:

$$F_{ij} = \frac{\sum_{k} LF_{ijk}}{0.186n_{ij}}$$
(1)

where LF_{ijk} is the number of live preemergent fry found in **dig** k, **stream** i, zone j, and n_{ij} is the number of digs from stream i, zone j. Densities of dead preemergent fry as well as dead and live embryos were found using the same estimator with appropriate substitutions.

Pink salmon overwinter survivals (\hat{S}_{ij}) were estimated as:

$$3_{ij} = \frac{\sum_{k}^{LF} LF_{ijk}}{\sum_{ijk} (LE_{eijk} + DE_{eijk} + LF_{eijk} + DF_{eijk})}$$
(2)

where LF_{fijk} is the number of live fry from dig k, stream i_i , zonej, collected during the fry survey, and n_e and n_f are the number of digs for stream i_i , zonej for the embryo and fry surveys.

Overwinter survival **data** were edited prior to analysis to remove values greater than 2.0, i.e., survivals greater than 200%.

Differences in overwinter survival (Y_{ijk}) were examined using a fixed effects, two factor experiment with repeated measures on one factor (Neter et al. 1990):

$$Y_{ijk} = \mu_{...} + O_i + Z_j + (OZ)_{ij} + S_{k(i)} + \varepsilon_{(ijk)} \qquad . \tag{3}$$

The two treatments were extent of oiling, (O_i) ; oiled and unoiled), and height in the intertidal zone (Z_j) ; 2.1, 2.7, and 3.4 m above mean low water and upstream). The **data** were blocked by stream $(S_{k(l)})$, which was nested within extent of oiling. The interaction of extent of oiling and height in the intertidal zone was also examined. The assumption of constant variance for error terms was tested using the F_{max} -test (Sokal and Rohlf 1969) while normality of error terms was visually assessed using scatter plots, box plots, and normal probability plots. Arcsin square root, logit, log, and square root transforms were examined if the **data** indicated non-constant variances or non-normal error terms. Assumptions relating to a valid split-plot analysis of the repeated measures factor, zone, were also examined. Tests of homogeneity of between-treatment covariance matrices and the degree of sphericity of the pooled covariance matrix (Mauchly 1940) were effected. Four contrasts (oil contaminated vs. control for the 4 stream zones) and corresponding Bonferroni family confidence intervals (a = 0.10 overall) were estimated if a significant difference due to oiling was detected. The SAS(Institute Inc. 1988) General Linear Models Procedure was used to analyze the **data**.

Embryo Rearing

We felt the stream effect could be removed and the genetic damage hypothesis evaluated through a designed experiment where embryos from *oil* contaminated and control streams were incubated in a common environment. Gametes from 30 male and 30 female pink salmon were collected from each of eight oil impacted and eight control streams in southwestern Prince William Sound (Figure 4) and flown to the Armin F. Koernig hatchery. Each oil contaminated stream was paired with a control stream based on similarity of geographic location and physical characteristics. Paired streams were sampled on the same day.

Gamete collection techniques were similar at each stream. Adults were captured at low tide in the stream mouth using a 30-m hand operated beach seine, and held in shallow water after being encircled. Only gametes from ripe individuals (adults that readily extruded eggs or sperm when gently massaged) were taken. Eggs from individual females (approx. 1500 per female) were removed by excising the abdominal wall, allowed to flow directly into 1-L ziplock plastic bags, and packed on cotton towels over a 10-cm layer of wet ice in insulated ice chests. Sperm from individual males (2-3 mls) was placed into 15-ml plastic centrifuge tubes, which were then capped and placed on ice in the same chests as the females for that stream. Once all gametes were collected on a stream, the ice chests were flown back to Armin F. Koernig Hatchery (an average 10 minute flight time) while gametes from the next stream were being collected.

The construction of stream specific embryo pools consisting of all single-pair crosses (30 x 30 = 900) was begun immediately upon arrival of the gametes at the hatchery. Crosses were made by first placing 5-ml of eggs (approximately 30 eggs) from each female into each of 30, 0.47 liter cups (each cup contained a teaspoon of eggs from each female). Each cup was then fertilized by a single male using 100 μ l of sperm followed by 100-ml of freshwater (8 °C) to initiate fertilization. This procedure provided each male an equal opportunity to fertilize eggs from each female. The fertilized eggs were allowed to sit for approximately 3-min after which they were recombined into a 3-L plastic container (maintained at **8** °C) and gently rinsed and mixed with freshwater three times.

Each day of stream sampling was placed into one of four stacks of Heath trays (FAL/Heath Tray, Tacoma, Washington, U.S.A.). Six trays within each of the four stacks were previously divided into 16 compartments (four rows by four columns) using plastic strips. Each strip was silicone sealed to the tray to prevent mixing of eggs and larvae between compartments. Twenty four samples of approximately 580 embryos (100-ml of embryos) each were randomly collected from the stream specific embryo pools and loaded into separate compartments using a prearranged random loading scheme. Only 18 samples were collected



Figure 4. Map of the study area in southwestern Prince William Sound, Alaska. Index number next to stream locator indicates the order of gamete collection.

from **4** of the streams and 12 samples from 2 streams on August 26, 1993 when 6 streams were sampled.

A technician was stationed at the hatchery during the three months of the experiment to perform normal fish culture duties and collect mortality data. The technician was made aware of the day of collection for record keeping but did not know which incubator compartments represented oiled or unoiled streams.

Dead eggs in each compartment were counted and removed at 36 hours after fertilization, after which trays were undisturbed for four weeks. Water flow to each of the four incubator stacks was maintained at four gpm. Each incubator stack received a 20-ppt sodium chloride bath for 20 minutes duration twice per week to control fungus infestations on the eggs. Water temperatures during incubation ranged from a high of 11°C to a low of 5 °C.

Mortality of eyed embryos (the point at which **a** distinct embryo eye could be seen through the chorion) was recorded at 350 temperature units (T.U.; $1 \text{ T.U.} = 1 \,^{\circ}\text{C}$ above $0 \,^{\circ}\text{C}$ for a 24-hr period). Embryos at this stage were siphoned out of their compartments using Tygon tubing (10 mm inside diameter) and allowed to drop 10-12 cm into a container of freshwater. The resulting physical shock caused coagulation of yolk material in undeveloped eggs allowing easier identification and removal. Live and dead eggs and embryos were gently placed back into their original compartments after siphoning. The live embryos were enumerated and the dead eggs were removed and counted.

Mortality was again recorded after the embryos had completely hatched (770 T.U.). In addition, the number of abnormal larvae (deformities of the head, body, or tal) in each compartment was also recorded. All larvae were destroyed after hatching.

The statistical difference in survival (Y) due to oil contamination (O)was evaluated using a blocked (day; D) analysis of variance:

$$Y_{ijk} = \mu_{..} + D_i + O_j + \varepsilon_{ijk} \qquad . \tag{4}$$

Flow Cytometry

Tissue Preparation

Flow cytometry was used to test for genetic damage (chromosome breakage) at several points in the project. Preparation of all tissue types followed similar procedures. Suspensions of stained nuclei were produced for **DNA** content analysis using nuclear isolation medium (NIM) (0.9% NaCl, 10 mM Tris, 2 mM CaCl₂, 2 mM MgCl₂, 0.1% Noniodet P-40, **106** mM MgSO₄, and 1 mg/100ml DAPI (4,6-diamidino-2-phenylindole dihydrochloride)). Fry, embryos and tissue samples were placed into 1.5 ml microcentrifuge tubes containing 1 ml of NIM. Samples were minced in 0.5 ml of NIM approximately one minute using two scalpels to obtain a cellular suspension, allowed to incubate at 2-3° C for 15 min, and filtered through a 40 μ m nitex nylon filter to remove debris and clumped cells. Stained nuclear suspensions were refrigerated overnight for flow cytometry analysis the following day. Immediately prior to flow cytometry analysis samples were triturated 3 times using a 26 g syringe and filtered through 40 μ m nitex nylon filter to remove any residual clumps of nuclei. Samples were analyzed using a PARTEC PAS II flow cytometer with optical filters for DAPI excitation and ACQCYTE data acquisition and MULTICYCLE DNA analysis software (Phoenix Flow Systems Inc. 1991) following the methods of Lamb et al. (1991).

Prince William Sound Preemergent Fry

Live fry from both oil contaminated and control **streams** were collected as part of the spring preemergent fry study and shipped to the ADF&G Genetics Laboratory in April, 1993. Approximately 100 fry each were received from three oiled and three unoiled streams. Whole fry were examined for chromosome damage using flow cytometry.

Saltwater Exposed Rainbow Trout

Polyploids and heteroploid mosaics were observed in November, 1992, during the initial flow cytometry analysis of pink salmon embryos from the 1992 Little Port Walter (LPW) oiling experiment. We were immediately interested in determining what factor caused the ploidy alterations in these embryos. Two controlled saltwater exposure experiments were conducted during April, 1993, to determine **if** saltwater exposure at fertilization could be responsible for inducing the ploidy alterations observed in the LPW pink salmon samples (Miller et. al. *In press*). As part of this experiment, an extensive screening of intertidally spawned pink salmon fry from Prince William Sound was performed to determine if ploidy alterations were a naturally occurring phenomenon associated with intertidal spawning, and possibly a causal agent of the embryo mortality observed in this region.

pink Salmon Controlled Oiling Experiment

pink salmon fry samples were received during June, 1993, from the controlled oiling experiment conducted at LPW. Fifty individuals from seven exposure groups were received. Individual tissues (liver, kidney, gill, muscle, blood, and spleen) from these fry were analyzed for evidence of chromosome damage.

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Crude **Cil** Water Soluble Fraction Experiment

An experiment was conducted during August and September, 1993, to determine the potential of the water soluble fraction of crude oil to induce chromosome damage in pink salmon embryos. This experiment was conducted in ADF&G's Genetics Laboratory.

A water soluble fraction (WSF) was used to test for potential clastogenic effects of the water soluble components of crude oil. WSF was prepared by adding 100-ml of North Slope crude oil (obtained from the **NMFS**, Auke Bay Laboratory's supply of Exxon Valdez crude oil) to 2 L of freshwater in a 2 L glass separatory funnel. The mixture was shaken at 8" C for 15 minutes, then allowed to separate in the funnel for 24 hr. The clear portion was drained into a glass stoppered bottle and used as a WSF stock solution (designated 100% WSF). A 1 L sample was sent to **an** independent environmental testing company for hydrocarbon analysis to identify the water soluble hydrocarbons present.

Glass petri dishes containing 30 ml of one of five concentrations of WSF (0%, 1%, 10%, 50%, and 100% WSF) were arranged in five groups, each group containing all five concentrations of WSF. **Pink** salmon gametes were obtained from 9 females and 9 males from the Armin F. Koernig hatchery on August 24, 1993, and **transported** by air back to the Genetics Laboratory. The experiment was begun the next day. Eggs and sperm were separately pooled prior to fertilization. Approximately 2500 eggs were fertilized with 200 μ l of sperm, activated with well water, and allowed to fertilize for one minute. Fifty ml of embryos (approximately 100 embryos) were randomly placed into the dishes within each of the five groups. This process was repeated three times.

Embryos were incubated in treatment water for 23 days at 7 °C prior to flow cytometry analysis. During this time, fresh WSF stock solution was generated and treatment water within the dishes was replaced every 36 hours.

Embryos were randomly chosen from each dish for flow cytometry analysis. Embryos were sampled after axis formation but prior to complete tissue differentiation (organ formation). Prepared embryos were coded and randomized prior to flow cytometry so that treatment was unknown during analysis.

RESULTS

Recovery Monitoring of Injury to Pink Salmon Eggs and Preemergent Fry in Prince William Sound

Mean preemergent fry densities for the 1993 fry survey were 220.30 fry per m^2 in the intertidal zones and 160.84 fry per m^2 in the upstream (Appendix A). The 1992 to 1993 embryo to preemergent fry survival information indicated no oil or oil by zone interaction (P=0.588, P=0.454; Figure 5). There was evidence for a zone effect (P=0.006) indicating that survivals were different by stream zone. The greatest survival was observed in the two highest intertidal zones.

Venjication of Injury to Pink Salmon Gametes in Prince William Sound

Gamete collection began on four streams on August 17, 1993. On that day, a large number of pink salmon were examined in order to obtain an adequate number of ripe fish; consequently, only two streams were sampled on the next day, August 18, 1993. Six streams were sampled on August 26, 1993, and four streams on August 27, 1993. Ripe fish were easily collected during the latter two days. The pink salmon embryos were shocked and picked during the period September 17-30, 1993 and the experiment was terminated on November 1, 1993. The summarized data collected from this experiment are presented in Appendix B.

Embryos from oil contaminated streams had lower survival at the eyed stage than those from control streams on three of the four days (Figure 6). Average eyed embryo survival was statistically different (P=0.012) between the control and oil contaminated streams (average survivals of 0.881 and 0.788, respectively). Survival at hatch was on the average 0.014 less than at eyeing for both the control and oil contaminated groups.



Figure 5. Mean pink salmon embryo to preemergent fry survival and corresponding 90% confidence bounds by tide zone for oil contaminated and control streams in Prince William Sound, 1993. Solid circles indicate oil contaminated streams while open circles identify control streams.



Figure 6. Mean pink salmon embryo survivals observed under hatchery conditions in 1993. Solid circles indicate oil contaminated streams while open circles identify control streams.

Flow Cytometry

The terms G1 and G2 will be used throughout the following section. G1 refers to cells in the normal diploid stage while G2 refers to cells near the completion of mitosis.

Prince William Sound Preemergent Fry

A total of X0 to X7 fry were analyzed using flow cytometry for each of three oil contaminated and three unaffected streams (Table 1). No significant difference in the coefficient of variation (CV) of the G1 peak was observed between fry from Contaminated and control streams (P = 0.668; one-way analysis of variance). Mean CV of the G1 peak from control streams was 4.36 (SD=0.202) and from control streams was 4.40 (SD=0.254).

Saltwater Exposed Rainbow Trout

Salt water was found to induce ploidy alterations in salmonid embryos (Miller et al. *In press*). Flow cytometry analysis revealed significantly higher frequencies of haploids, triploids, heteroploid mosaics, and aneuploids in rainbow trout embryos experimentally exposed to saltwater from fertilization to the two and eight-cell stages of development. Heteroploid mosaics have been reported in diploid and triploid salmonid hybrids; although, none were observed in the triploid coho salmon or diploid and triploid coho salmon X chinook salmon hybrids we examined. No mosaics were observed in intertidally spawned pink salmon embryos. Heteroploid embryos appeared morphologically normal, although, they may possess physiological deficiencies not immediately apparent.

Pink Salmon Controlled Oiling Experiment

No significant differences were observed in the mean CV of the G1 peak or in the mean G1/G2 ratio of cells between the treatment groups in the tissues analyzed from the brood year 1992 fry from the Little Port Walter controlled oiling experiment (Table 2). No data for blood G1/G2 ratio are presented since fish erythrocytes are mature cells and do not possess a G2 phase of the cell cycle.

Crude Oil Water Soluble Fraction Experiment

No differences in the coefficient of variation were observed in the WSF exposure groups (Table 3). There was an indication of a decrease in the G1/G2 ratio with increasing oil exposure, although, this difference was not statistically significant (P=0.0545; one-way analysis of variance on natural log transformed data). The percentage of eggs containing viable embryos appeared to decrease with increasing WSF exposure (Table 4); although, we were unable to test this trend due to sampling limitations.

Table 1.Mean coefficient of variation (CV) for the G1 cell cycle, standard deviation
(SD), and sample size (n) for pink salmon fry from three oil contaminated and
three unaffected streams in Prince William Sound, Alaska. Whole fry were
analyzed using flow cytometry.

		Mean		
Stream	Oil	CV G1	S.D.	n
604	No	4.24	0.83	80
630	No	4.59	1.02	80
695	No	4.24	0.66	87
618	Yes	4.52	0.83	80
637	Yes	4.16	0.53	80
682	Yes	4.65	0.72	87

Table 2. Summary of flow cytometry results from the Little Port Walter controlled oiling experiment. Data are from analysis of individual tissues from brood year 1992 oil exposed pink salmon fry. The coefficient of variation (CV) of the G1 peak and the G1/G2 ratio of cells are presented for each exposure group along with the corresponding standard deviation (SD). Significant differences between oil exposure groups were tested by one-way analysis of variance on natural log transformed data.

Tissue	Oil Exposure (g oil/kg substrate)	n	Mean CV	SD	Р	Mean G1/G2	SD	Р
Blood	0.0	13	3.88	0.50	0.924			
	0.1	12	4.26	0.90				
	0.4	10	4.24	0.94				
	1.5	13	4.02	0.84				
	5.7	12	3.98	0.88				
Gill	0.0	10	4.62	2.67	0.145	32.72	21.81	0.687
	0.1	8	7.01	5.72		24.13	14.19	
	0.5	7	3.47	1.52		31.90	26.58	
	1.5	7	4.29	1.60		27.00	23.26	
	5.7	7	8.54	7.53		38.24	24.28	
Kidney	0.0	4	2.73	0.61	0.171	20.74	15.29	0.507
	0.1	3	3.90	2.38		15.58	4.04	
	0.5	4	2.98	0.90		34.61	5.95	
	1.5	3	4.31	1.69		30.64	31.80	
	5.7	5	2.28	0.22		51.46	37.69	
Liver	0.0	12	2.76	0.51	0.171	84.98	62.01	0.973
	0. I	12	2.35	0.42		76.24	45.52	
	0.5	12	2.77	0.63		67.16	38.48	
	1.5	13	2.45	0.40		75.03	55.12	
	5.7	10	2.71	0.29		68.28	67.02	

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Tissue	Oil Exposure (g oil /kg substrate)	n	Mean CV	SD	Р	Mean G1/G2	SD	Р
Muscle	0.0	11	3.65	0.49	0.649	22.22	7.10	0.299
	0.1	9	3.90	0.52		16.35	5.37	
	0.5	9	4.00	0.78		16.39	5.90	
	1.5	12	3.98	0.58		19.75	13.34	
_	5.7	8	3.81	0.42		18.14	7.67	
Spleen	0.0	12	2.98	1.04	0.961	52.04	32.18	0.334
	0.1	11	3.25	0.88		41.35	18.36	
	0.5	10	3.35	1.25		30.12	25.23	
	1.5	12	2.99	0.38		51.97	25.08	
	5.7	11	3.16	0.67		51.78	28.13	

WSF			Mean	Cell Statisti	cs and Standa	rd Errors	
Exposure	n	CV GI	% Cells G1	CV G2	% Cells G2	% S	G1/G2ª
0 %	29	3.2 (0.12)	62.0 (0.95)	3.5 (0.16)	7.4 (0.38)	30.6 (1.01)	10.1 (1.36)
1%	28	3.3 (0.08)	64.5 (0.96)	3.8 (0.12)	7.0 (0.39)	28.5 (0.84)	11.1 (1.53)
10 %	25	3.1 (0.07)	63.0 (0.82)	3.5 (0.11)	8.1 (0.30)	28.9 (0.69)	8.1 (0.40)
50 %	29	3.4 (0.16)	61.4 (0.82)	3.8 (0.19)	8.1 (0.46)	30.5 (0.89)	8.8 (0.82)
100 %	29	3.2 (0.09)	61.8 (1.01)	3.6 (0.12)	8.4 (0.30)	29.8 (0.81)	7.7 (0.41)

Table 3.Flow cytometry analysis results from **pink** salmon embryo exposure to *Exxon*
Valdez water soluble fraction (WSF). Mean cell cycle statistics are presented
with corresponding standard errors in parentheses.

^a P=0.054 for a test of equality of means.

		Percent	Eggs
WSF Exposure	n	No Embryo	Embryo
0%	101	59.4 (60)	40.6 (41)
1%	116	64.7 (75)	35.3 (41)
10%	107	72.9 (78)	27.1 (29)
50%	105	63.8 (67)	36.2 (38)
100%	170	76.5 (130)	23.5 (40)

Table 4.Percentage of pink salmon eggs with and without embryos at the time of
sampling for flow cytometry. Actual number observed are in parentheses.

DISCUSSION

Pink salmon embryos which incubated in oil contaminated spawning areas in Prince William Sound appear to have been adversely affected by the *Exxon Valdez* oil spill. Sharr et al. (1994a and 1994b) found increased pink salmon embryo mortalities in 1989, 1990, 1991, and 1992. We believe the elevated mortalities observed in 1989 and 1990 were due to direct exposure to oil while those in 1991 and 1992 are hypothesized to be due to genetic damage sustained during embryonic and alevin development.

The pink salmon which spawned during the fall of 1991 were the fry which incubated in oil contaminated streams during winter of 1989-1990, the first winter after the spill. Likewise, the pink salmon which spawned during the fall of 1992 were the same fry which incubated in oiled stream gravel during the fall of 1990 and spring of 1991. Sharr et al. (1994a) found significantly elevated embryo mortalities in oil contaminated streams during the fall of 1989 and 1990, and there is a strong possibility the surviving embryos sustained sublethal genetic damages which were manifested in the form of functional sterility in 1991 and 1992.

The alternative to the genetic damage hypothesis is that observed differences in embryo mortality are due to environmental variation. The embryo mortality study is based on observational data, and as such, we were unable to randomize stream oiling to account for environmental differences between streams. We attempted to address this concern in our original experimental design by selecting unoiled or control streams in close proximity to oil contaminated streams; however, there is a definite oiling pattern in southwest Prince William Sound where streams on points which faced northeastward were heavily oiled. Likewise, streams which faced west and southwest were most likely not oiled.

We found strong evidence to suggest that the basis for the differences in embryo mortality observed in the field was carried by the parents. Six of the eight oil contaminated streams sampled had lower embryo survival than the uncontaminated streams (Figure 6). The gametes used in this study had never been in contact with a stream; although, the adults which produced them had incubated in the natal streams. This work examined the 1993 brood year which was removed by the oil spill by two generations (1989 and 1991 broods).

Although flow cytometry analysis of oil exposed whole embryos and individual tissues has not yet detected significant levels of chromosome damage, differences in the timing of the cell cycle were apparent. The decreasing G1/G2 ratio with increasing oil exposure (Table 3) indicates cell division is being delayed immediately prior to the completion of mitosis. This delay could be caused by chromosome or DNA damage interfering with **DNA** replication or cell division, by toxicologic effects of crude oil delaying cell division, or both.

Again this data does not provide definitive evidence that the differences are caused by an oil contaminated environment. It is conceivable; although, we don't believe probable, that the streams which were oiled have also historically had lower pink salmon embryo survival rates.

The controlled oiling experiment portion of this work being done by the National Marine Fisheries Service will provide laboratory evidence to support or refute this field data.

No difference in embryo to preemergent fry survival has been detected. We expected embryo to preemergent *fry* survival to be reduced in oil contaminated streams for all years examined given that an increase in embryo mortality was already detected; this has not been the case. We suspect that unexpected changes in stream characteristics prevented us from sampling the same areas or populations for embryos in the fall and *fry* in the spring. Runoff from fall rains increase stream depth and width while spring water levels are usually low since the majority of the winters precipitation is tied up in ice and snow. Also stream channels in Prince William Sound are not well defined in intertidal areas. It is common for intertidal stream segments to migrate along the beach especially if the beach is exposed to winter storms. The magnitude of these changes was unexpected when this study **was** designed and initiated.

CONCLUSIONS

There is strong evidence to suggest that the increased pink salmon embryo mortalities observed in oil contaminated streams during the falls of 1991 and 1992 were the result of traits carried by their parents.

No difference in pink salmon embryo to preemergent fry survival were observed during the winter and spring of 1992-1993.

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					Pin< Salmon						Chum Salmon						
Street	Streen				E	ggs	Ē	Γ γ	Live	Fry/m ²	Eg	gs	Fr	'Y	Live	Fry/m²	
# 	Name	Date	Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
11	Humpy Creek	3 12 93	2.13 2.74 3.35 Upstream Upstream	20 30 40 61	0 0 6 0 0	00000	0 0 7 0	0 0 35 0 0	.00 .00 13.44 .00 .00	00 00 11.88 00 00	0 0 0 0	0 0 0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14 14
		Tota Tota	l Intertidal l Upstream		6 0	0 0	7 0	35 0	4.59 .00	4.08 .00	0 0	0 0	0 0	0 0	.00 .00	.00 .00	42 24
35	Koppen Creek	3 15 93	2.74 3.35 Upstream	30 40 60	31 120 648	2 1 0	2 2 2	1262 3216 1	484.52 1234.71 .38	178.98 284.98 .38	0 0 0	0 0 0	0 0 0	0 2 0	.00 .77 .00	.00 .52 .00	14 14 14
		Total Total	l Intertidal l Upstream		151 648	3 0	5	4478 1	859.62 .38	180.21 .38	0 0	0 0	0 0	2 0	.38	.27	28 14
52	Control Creek	3 16 93	2.13 2.74 3.35 Upstream	20 30 40 60	117 266 636 211	0 0 0 0		380 488 344 101	145.89 187.36 132.07 38.78	99.21 98.50 52.87 27.21	0 0 0	0 0 0	0 0 0	0 3 0 2	.00 1.15 .00 .77	.00 1.15 .00 .77	14 14 14 14
		Total Total	. Intertidal . Upstream		1019 211	0 0	2	1212 101	155.11 38.78	48.73 27.21	0 0	0 0	0 0	3 2	.38 .77	-38 -77	42 14
80	Whalen Creek	4 6 93	2.13 2.74 3.35 Upstream	20 30 40 60	20 32 1041 229	0 0 0 0	0 0 3 0	14 259 235 29	5.38 99.44 90.22 11.13	2.25 46.53 52.92 6.31	0 0 6 2	0 0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
		Total Total	Intertidal Upstream		1093 229	0 0	3 0	508 29	65.01 11.13	23.86 6.31	6 2	0 0	0 0	0 0	.00 .00	.00	42 14

Appendix A. 1993 Prince William Sound Pink and Chum Salmon Fry Dig.

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							Pink	Salmor	n				Chur	n Salmo	on		
					Eg	jgs	F	гу	Live	₽ Fry/m²	Eg	gs	Fr	·у	Live	Fry/m ²	
Stream #	Stream Name	D⊧te	Height in Tidal Zone(m)	Lo≺	- Dead	Live	Dead	Live	Mean	ŞE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
89	Fish Creek	3 17 93	2.13 2.74 3.3 5 Upstream	20 30 40 60	0 22 364 479	0 0 0 0	0 0 0 0	7 1650 166 0	2.69 633.48 63.73 .00	1.85 195.34 35.44 .00	0 1 63 0	0000	0 0 0 0	0 6 25 0	.00 2.30 9.60 .00	.00 1.09 9.60 .00	14 14 14 14
		Tota Tota	al Intertidal al Upstream		386 479	0 0	0 0	182 3 0	233.30 .00	78.32 .00	64 0	0 0	0 0	31 0	3.97 .00	3.20 .00	42 14
117	Indian Creek	3 18 93	2.13 2.74 3.35 Upstream	20 30 40 60	117 122 55 16	0 0 0 0	0 13 0 1	483 910 1539 793	185.44 349.38 590.87 304.46	58.35 107.22 236.61 151.79	0 0 23 5	0000	0 0 0 0	1 0 646 1230	.38 .00 248.02 472.23	.38 .00 113.30 262.62	14 14 14 14
		Tota Tota	l Intertidal l Upstream		294 16	0 0	13 1	2932 79 3	375.23 304.46	90.38 151.79	23 5	0 0	0 0	647 1230	82.80 472.23	41.10 262.62	42 14
123	Gregorieff Creek	3 18 93	2.13 2.74 3.35 Upstream	21 31 41 61	0 55 344 222	0 11 0 25	0 1 27 24	2 1 1256 1641	.77 .38 482.21 630.03	.52 .38 187.53 219.21	0 0 0 0	0000	0 0 0 0	0 0 24 25	.00 .00 9.21 9.60	.00 .00 5.67 8.03	14 14 14 14
		Tota Tota	l Intertidal l Upstream		399 222	11 25	28 24	1259 1641	161.12 630.03	70.53 219.21	0 0	0 0	0 0	24 25	3.07 9.60	1.97 8.03	42 14
153	Stellar Creek	4793	2.13 2.74 3.35 3.35 Upstream	20 30 40 43 60	45 480 151 8 0	0 0 0 0	0 0 17 30 0	158 778 1154 251 6	60.66 298.70 443.05 96.37 2.30	37.67 89.76 126.28 32.63 .74	0 0 1 52 2	00000	0 0 1 177 0	0 60 2121 10	.00 .00 23.04 814.31 3.84	.00 .00 19.39 341.93 2.84	14 14 14 14 14
		Tota Tota	l Intertidal l Upstream		684 0	0 0	47 0	2341 6	224.69 2.30	44.77 .74	53 2	0	178 0	2181 10	209.34 3.84	95.66 2.84	56 14

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					Egg]s	Fr	у	Live	Fry/m²	Eg	igs	Fr	у	Live	Fry/m ^z	
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
265	Unakwik Creek	3 19 93	2.13 2.74 2.74 3.35 3.35 Upstream Upstream	20 31 32 41 42 61 62	0 80 0 0 0	0 0 0 0 0	0 0 1 0 0 0	406 733 1113 0 21 0 0	155.88 562.84 854.63 .00 16.13 .00 .00	88.81 208.68 165.50 .00 9.24 .00 .00	000000	0000000	0 0 0 0 0 0	0 0 0 0 0 0	.00 .00 .00 .00 .00 .00	.00 .00 .00 .00 .00 .00	14 7 7 7 0 0
		Tota Tota	l Intertidal l Upstream		80 0	0 0	1 0	2273 0	290.89 .00	70.41 .00	0 0	0	0	0 0	.00 .00	.00 .00	42 0
276	Black Bear Creek	3 19 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 26 166 116	0 0 0 0	0 0 14	0 103 1086 641	.00 39.54 416.95 246.10	.00 32.75 132.50 164.67	00000	0000	0 0 0 0	0 35 0 246	.00 13.44 .00 94.45	.00 13.03 .00 51.39	14 14 14 14
		Tota Tota	l Intertidal l Upstream		192 116	0 0	0 14	1189 641	152.16 246.10	53.20 164.67	0 0	0	0	35 246	4.48 94.45	4.35 51.39	42 14
3 22	Coghill River	3 20 93	Upstream	60	245	0	0	0	.00	.00	0	· C	0	0	.00	.00	60
		Tota Tota	l Intertidal l Upstream		0 245	0 0	0 0	0 0	.00 .00	.00 .00	0 0		0	0 0	.00 .00	.00 .00	0 60
421	Mill Creek	3 20 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 4 10 0	0 0 0 0	0 3 0 0	0 590 209 0	00 226.52 80.24 .00	.00 162.02 44.87 .00	0 31 0 0		0 1 0 0 0	0 68 29 0	.00 26.11 11.13 .00	.00 18.44 5.09 .00	14 14 14 14
		Tota Tota	l Intertidal l Upstream		14 0	0 0	3 0	799 0	102.25 .00	56.58 .00	31 0) () 1) 0	97 0	12.41 .00	6.44 .00	42 14

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							Pink	Salmo	n				Chu	n Salmo	on		
Stream	Streem				Eg	gs	F	ry	Liv	e Fry/m²	Eg	gs	Fr		Liv	e Fry/m²	
#	Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mear	n SE	Dead	Live	Dead	Lîve	Mear		No. of
430	Meacham Creek	3 21 93	2.13 2.74 3.35 Upstream	20 30 40 60	61 312 4 138	0 0 0 0	0 0 0 1	0 56 373 726	.00 21.50 143.21 278.73	.00 13.50 94.51 112.65	0 0 6 0	0 0 0	0 0	0 0 64	.00 .00 24.57	.00 .00 21.39	14 14 14 14
		Tota Tota	l Intertidal Upstream		377 138	0 0	0 1	429 726	54.90 278.73	32.56 112.65	6 0	0 0	0	64 0	8.19 .00	.00 7.19 .00	14 42 14
455	Paulson Creek	3 21 93	2.13 2.74 3.35 3.35 Upstream	20 30 40 43 60	13 1 0 17	0 0 0 0	0 0 0 13	0 486 37 125 631	.00 186.59 28.41 95.98 242.26	.00 77.07 17.50 82.09 96.34	0 2 20 3 0	00000	0 0 0 0 0	1 141 0 2	.38 54.13 .00 .00 77	.38 16.42 .00 .00	14 14 7 7
		Total Total	Intertidal Upstream		14 17	0	0 13	648 631	82.93 242.26	30.91 96.34	25 V	0 0	0	142 2	18.17 .77	6.66	42
480 1	1ink Creek	3 22 93	2.13 2.74 3.35 Upstream	20 30 40 60	68 23 1 45	0 0 0 0	1 0 7 0	595 1063 1411 1	228.44 408.12 541.72 .38	93.85 143.40 157.23 .38	0 0 2 0	0 0 0	0 0 0	255 9 45	97.90 3.46 17.28	91.42 3.46 14.92	14 14 14
		Total Total	Intertidal Upstream		92 45	0 0	8 0	3069 1	392.76 .38	78.22	2 0	0 D	0	309 0	.00 39.54 .00	.00 30.83 .00	14 42 14
485 W	l. Finger Creek	4 7 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 62 15 16	0 0 0 0	0 22 1 2	216 939 1378 874	82.93 360.51 529.05 335.55	51.78 180.58 239.55 184.33	0 0 0 0	0 0 0 0	0 0 0	0 27 4 0	.00 10.37 1.54 .00	.00 4.89 1.54 .00	14 14 14 14
		Total Total	Intertidal Upstream		77 16	0 0	23 2	2533 874	324.16 335.55	103.05 184.33	0 0	0 0	0 0	31 0	3.97	1.81	42

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								Pink	Salmor	ı				Chur	n Salmor	n		
						Eg	jgs	Fr	у	Live	Fry/m ²	Eg	gs	Fr	у	Live	Fry/m ²	
Stream #	Stream Name	Da	ate	Height in Iidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	 Dead	Live	Mean	SE	No. of Samples
498	McClure Creek	4 1	3 93	2.13 2.74 3.35 Upstream	20 30 40 60	2 177 14 22	0000	0000	364 2556 2900 0	139.75 981.32 1113.39 .00	51.55 232.77 428.39 .00	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
			Total Total	Intertidal Upstream		193 22	0 0	0 0	5820 0	744.82 00,	173.03 .00	0 0	0 0	0 0	0 0	.00 .00	.00 .00	42 14
506	Loomis Creek	4	793	2.13 2.74 3.35 Upstream	20 30 40 60	39 834 919 1008	0000	0 1 0 0	721 1026 276 362	276.81 393.91 105.96 138.98	145.83 99.89 51.66 77.05	0 4 0 0	0 0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
			Total Total	Intertidal Upstream		1792 1008	0 0	1 0	2023 362	258,90 138,98	62.65 77.05	4 0	0 0	0 0	0 0	.00 .00	.00 .00	42 14
604	Erb Creek	4	893	2.13 2.13 2.74 3.35 Upstream	20 23 30 40 60	35 28 140 224 5	00000	1 0 0 36 0	186 192 569 398 0	142.82 147.43 218.46 152.80 .00	63.62 67.06 106.38 83.57 .00	0 0 0 0	00000	0 0 0 0	0 0 39 0 0	.00 .00 14.97 .00 .00	.00 .00 13.78 .00 .00	7 7 14 14 14
			Total Total	Intertidal Upstream		427 5	0 Q	37 0	1345 0	172.13 .00	46.57 .00	0 0	0 0	0 0	39 0	4.99 .00	4.61 .00	42 14
618	Junction Creek	4	293	2.13 2.74 3.35 Upstream	20 30 40 60	0 0 11 1	0000	0 0 0	0 0 380 0	.00 .00 170.21 .00	.00 .00 170.21 .00	0 0 0 0	0000	0 0 0 0	0 0 0 0	.00 .00 .00	.00 .00 .00 .00	12 12 12 12
			Total Total	Intertidal Upstream		11 1	0	0 0	380 0	56.74 .00	56.74 .00	0 0	00	0 0	0 0	.00	.00 .00	36 12

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								Pink	Salmor	L				Chur	n Salmo	n		
						Eg]gs	Fi	у	Live	Fry/m ²	Eg	gs	Fr	у	Live	e Fry/m²	
Stream #	Stream Name	C	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mear	n SE	No. of Samples
621	Totemoff Creek	4	893	2.13 2.74 3.35 Upstream	20 30 40 60	138 19 320 474	0 0 0 0	0 0 33 1	453 607 2321 0	173.92 233.04 891.10 .00	82.56 87.28 221.31 .00	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	8666	0000 0000	14 14 14 14
			Tota Tota	l Intertidal l Upstream		477 474	0 0	33 1	33 81 0	432.69 .00	96.33 .00	0 0	0 0	0 0	0 0	90 90	.00 .00	42 14
623	Brizgaloff Creek	4	893	2.13 2.74 3.35 Upstream	20 30 40 60	58 282 2115 1274	0 0 0	0 0 23	345 747 1389 2565	132.46 286.79 533.28 984.78	59.62 124.98 337.62 187.13	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	6666 6666	0000 0000	14 14 14 14
			Tota Tota	l Intertidal l Upstream		2455 1274	0 0	0 23	2481 2565	317.51 984.78	121.40 187.13	0 0	0 0	0 0	0 0	00 00	95 95	42 14
628	Chenega Creek	4	2 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 20 9 16	0 0 4 0	0 1 0 0	0 585 73 0	.00 224.60 28.03 .00	.00 122.67 10.65 .00	0 0 0 0	0 0 0 0	0 0 0	0 0 0 0	8666	0000 00000	14 14 14 14
			Tota Tota	l Intertidal l Upstream		29 16	4 0	1 0	658 0	84.21 .00	42.97 .00	0 0	0 0	0 0	0 0	00 00	90 90	42 14
630	Bainbridge Creek	4	993	2.13 2.74 3.35 Upstream	20 30 40 60	58 419 9 263	0 0 0 0	0 0 3 1	89 1734 1901 2020	34.17 665.73 345.92 775.54	18.59 195.51 143.36 241.45	0 0 0	0 0 0 0	0 0 0 1	0 0 ZP4 0	.00 .00 112.88 .00	.00 .00 86.49 .00	14 14 14 14
	x		Tota Tota	l Intertidal Upstream		486 263	0 0	3 1	2724 2020	348.61 775.54	88.72 241.45	0 0	0 0	0 1	Z94 0	37.63 .00	zبر ²⁹ 00	42 14

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							Pink	Salmor	ı				Chur	n Salmo	n		
					Eg	jgs	Fr	y	Live	Fr y/m²	Eg	gs	Fr	y .	Live	Fry/m ²	
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
632	Claw Creek	4 9 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 10 166 133	0 0 0 0	0 0 0 0	2 3 1295 0	.77. 1.15 497.19 .00	.52 1.15 131.37 .00	0 1 0	0 0 0		0 0 0	.00 .00 .00	.00 .00 .00	14 14 14
		Tot Tot	al Intertidal al Upstream		176 133	0	0	1300 0	166.37 .00	56.20	1 0	0		0	.00 .00	.00 00	42 7
637	Pť. Countess	4993	2.13 2.74 3.35 3.35 Upstream Upstream	20 30 41 42 61 62	0 1 146 86 71 10	0 0 0 0 0	0 0 0 0 0	57 1006 119 1306 30 0	21.88 386.23 91.38 1002.82 23.04 .00	8.74 147.62 71.09 396.96 19.55 .00	0 0 0 0 0	0 0 0 0 0 0		0 0 0 0 0	00. 00. 00. 00. 00. 00.	.00 .00 .00 .00 .00 .00	14 14 7 7 7 7
		Tot Tot	al Intertidal al Upstream		233 81	0 0	0 0	2488 30	318.40 11.52	547 549 549	0 0	0 0	0 0	0 0	.00 .00	.00 .00	42 14
653	Hogg Creek	4 10 93	2.13 2.74 2.74 3.35 Upstream	20 31 32 40 60	19 1 0 1 161	0 0 0 0	4 0 0 0	41 168 1 27 44	15.74 129.00 .77 10.37 16.89	10.95 123.68 .77 9.96 13.68	0 0 0 0	0 0 0 0	0 4 0 0	0 52 0 0	.00 39.93 .00 .00	.00 39.93 .00 .00	14 7 7 14 14
		Tot Tot	al Intertidal al Upstream		21 161	0 0	4 0	237 44	30.33 16.89	21.08 13.68	0 0	0 0	4 0	5z 0	6·\\5 ·\\0	645 00	42 14

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							Pink	Salmo	'n				Chu	m Salmon	n		
Strea	n Stream		Vojaht iz		E	99s	F	гу	Live	e Fry∕m²	Eg	lgs	Fr	у	Live	Fry/m ²	
#	Name	Date	Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
656	Halverson Creek	4 10 93	2.13 2.74 3.35 Upstream	20 30 40 60	23 8 262 1218	0 0 0	0 1 1 72	758 664 4376 3238	291.02 254.93 1680.07 1243.16	193.46 114.43 324.30 273.77	0 0 0 0	0 0 0	0 0 0	0 0 0	.00	.00	14 14 14
		Tota Tota	l Intertidal l Upstream		293 1218	0 0	2 72	5798 3238	742.01 1243.16	164.90 273.77	0 0	0	0	0	.00 .00	.00 .00	14 42 14
663	Shelter Bay	3 30 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 0 178 68	0 0 0	0 0 0 0	0 8 103 0	.00 3.58 46.14 .00	.00 3.58 16.59 .00	0 0 0	0 0 0	000	0	.00 .00 .00	.00 .00 .00	12 12 12
		Tota Tota	l Intertidal l Upstream		178 68	0 0	0 0	111 0	16.57 .00	6.54 .00	0	0	0	0	.00 .00 .00	.00 .00 .00	12 36 12
665	Bjorne Creek	3 30 93	2.13 2.74 3.35 Upstream	20 30 40 60	27 3 474 232	0 0 0	0 0 115 2	0 66 293 0	.00 25.34 112.49 .00	.00 18.89 100.33 .00	0 0 0	0 0 0	0 0 0	0 0	.00 .00 .00	.00 .00 .00	14 14 14
		Total Total	Intertidal Upstream		504 23 2	0 0	115 2	359 0	45.94 .00	34.03	0	0	0	0	.00 .00 .00	.00 .00 .00	14 42 14
666	O'Brien Creek	3 31 93 Toto'	2.13 2.74 3.35 Upstream	20 30 40 60	0 0 42 59	0 0 0	0 0 1 0	0 198 17 0	00. 76.02 6.53 .00	.00 59.01 6.53 .00	0 0 0 0	0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
		Total	Upstream		42 59	0 0	1 0	215 0	27.51 .00	20.04 .00	0 0	0 0	0 0	0 0	.00 .00	.00	42 14

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					••		РІпк	Salmo	n				Chu	n Salmor	ו		
{tream	Stream		Voight in		E	9 9 \$	F	ry	Live	₽ Fr y/m ²	Eg	igs	Fr	у	Live	F-y/m²	
#	Name	Date	Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
673	Falls Creek	4 1 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 32 86 160	0 0 0 0	0 0 0 1	0 54 761 23	.00 20.73 292.17 8.83	.00 10.05 137.18 8.83	000000000000000000000000000000000000000	0 0 0	0 0 0	0 0 0	.00	.00 .00 .00	14 14 14
		Tot Tot	al Intertidal al Upstream		118 160	0 0	0 1	815 23	104.30 8.83	49.31 8.83	0	0	0	0	.00 .00	- 00 - 00 - 00	14 42 14
D77	Kayden Creek	4 1 µ3	2.13 2.13 2.74 2.74 3.35 3.35 Upstream Upstream	21 22 31 32 41 42 61 62	0 39 11 11 75 12 0 0	00000040		0 567 400 16 1 673 0 0	.00 435.38 307.14 12.29 .77 516.77 .00 .00	.00 181.47 178.50 5.83 .77 229.88 .00	0 0 0 0 0 0		000000000000000000000000000000000000000	000000000000000000000000000000000000000	99999999999999999999999999999999999999	.00 .00 .00 .00 .00 .00	7 7 7 7 7 7
		Tota Tota	al Intertidal al Upstream		148 0	0 4	0 0	1657 0	212.06 .00	63.35 .00	0 0	0	0 0	0	90 90 90	.00 .00	42 14
6'8 <u>9</u>	Sleepy Bay	3 30 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 21 15 31	0 0 0 0	0 0 0 0	0 214 512 0	.00 95.85 229.33 .00	.00 52.33 136.25 .00	0 0 0	0 0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	12 12 12 12
		Tota Tota	l Intertidal l Upstream		36 31	0 0	0 0	726 0	108.40 .00	49.84 .00	0 0	0 0	0 0	0 0	.00	.00 .00	36 12

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								Pink	Salmo	n				Chur	n Salmon			
						Eç	jgs	F	гy	Live	Fry/m ²	Eg	gs	Fr	y	Live I	Fry/m²	
Stream #	Stream Name		Date	Height in Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
681		3	29 93	2.13 2.74 3.35 Upstream	20 30 40 60	12 32 1941 1531	0 0 0 0	0 0 0 1	255 3 0 0	97.90 1.15 .00 .00	69.51 1.15 .00 .00	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
			Total Total	Intertidal Upstream		1985 1531	0 0	0 1	258 0	33.02 .00	23.71 .00	0 0	0 0	0 0	0 0	.00 .00	.00 .00	42 14
682		3	29 93	2.13 2.74 3.35 Upstream	20 30 40 60	234 216 456 395	0 0 0	0 0 3 43	947 3240 3443 5630	363,58 1243,93 1321,87 2161,52	130.24 222.24 585.71 673.22	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
			Total Total	Intertidal Upstream		906 395	0 0	3 43	7630 5630	976.46 2161.52	218.81 673.22	0 0	0 0	0 0	0 0	.00 .00	.00	42 14
692		4	693	2.13 2.74 3.35 Upstream	20 30 40 60	18 7 7 42	0 0 0 0	0 0 0 0	200 400 55 3	76.79 153.57 21.12 1.15	68.53 70.87 17.19 .83	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
			Total Total	Intertîdal Upstream		32 42	0 0	0 0	655 3	83.82 1.15	33.62 .83	0 0	0 0	0 0	0 0	.00 .00	.00 .00	42 14
695		4	2 93	2.13 2.13 2.74 3.35 Upstream	21 22 30 40 60	0 6 111 104 262	0 0 0 0	0 0 10 1	0 487 1985 1182 409	.00 373.95 762.10 453.80 157.03	.00 137.31 227.74 154.70 82.59	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00 .00	.00 .00 .00 .00	7 7 14 14 14
			Total Total	Intertidal Upstream		221 262	0 0	10 1	3654 409	467.63 157.03	100.51 82.59	0 0	0 0	0 0	0 0	-00 -00	.00 .00	42 14

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							Pink	Salmon					Chur	n Salmor			
Strea	m Stream				- Eg	jgs	F	гу	Live	e Fry/m²	Eg	gs	 fr	у	Live	Fry/m²	
#	Name	Date	Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
699	Cathead Bay	4 193	2.13 2.74 3.35 Upstream	20 30 40 60	140 14 274 436	0 0 0	0 0 0 12	599 1340 13 481	229.97 514.46 4.99 184.67	125.13 239.70 4.99 69.11	0 0 0 0	0 0 0	0 0 0	0 0 0	.00 .00 .00	.00.00	14 14 14
		Total Total	. Intertidal Upstream		428 436	0 0	0 12	1952 481	249.81 184.67	93.75 69.11	0 0	0 0	0	0	.00 .00	.00 .00	42 14
740	Kelez Creek	3 27 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 6 196 0	0 0 0 0	0 0 21 0	0 0 19 0	.00 .00 7.29 .00	.00 .00 7.29 .00	0 0 0 0	0 0 0	0 0 0	0 0 0	.00 .00 .00	.00 .00 .00	14 14 14
		Total Total	Intertidal Upstream		202 0	0 0	21 0	19 0	2.43 .00	2.43 .00	0 0	0 0	0	0	.00	.00 .00	42 14
744	Wilby Creek	3 28 93	2.13 2.74 3.35 Upstream	20 31 40 60	4 0 0 1	0 0 0	0 0 0	1 3 392 0	.38 1.15 150.50 .00	.38 1.15 139.53 .00	0 0 0 0	0 0 0	0 0 0	0 0 0	.00 .00 .00	.00 .00 .00	14 14 14
		Total Total	Intertidal Upstream		4 1	0 0	0 0	396 0	50.68 .00	46.68	0 0	0 0	0	0	.00	.00 .00	42 14
747	Cabin Creek	3 \$8 93	2.13 2.74 3.35 Upstream	20 30 40 60	0 122 391 223	0 0 0 0	0 0 183 0	1 25 1004 272	.38 9.60 385.46 104.43	.38 5.81 108.22 71.36	0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	-00 -00 -00 -00	.00 .00 .00 .00	14 14 14 14
		Total Total	Intertidal Upstream		513 223	0 0	183 0	1030 272	131.82 104.43	45.01 71.36	0 0) 0	0 0	5 3	.00	.00	42 14

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							Pink	Salmon	I				Chur	n Salmor	n		
					Eg	Igs	Fr	y	Live	Fry/m ²	Egg	js	Fr	Y	Live	Fry/m²	
Stream #	Stream Name	D∋te T	Height in idal Zone(m)	Ļс	Dead	Live	Dead	Live	- Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
749	Shad Creek	3 22 93	2.13 2.74 3.35 Upstream	20 30 40 60	43 6 0 216	0 0 0 0	0 0 4	174 20 9 644	66.80 7.68 3.46 247.25	48.49 7.68 1.92 147.62	0 0 0 0	0 0 0	0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
		Total Total	Intertidal Upstream		49 216	0 0	0 4	203 644	25.98 247.25	16.60 147.62	0 0	0	i 0 i 0	0 0	.00 .00	.00 .00	42 14
775	Pautzke Creek	3 23 93	2.13 2.74 3.35 Upstream	20 30 40 60	1 0 15	0 0	0 0	0 151 100 28	1 .38 57.97 38.39 10.75	54.38 54.36 37.16 6.83	(0 0 0		0) 0) 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	14 14 14 14
		Total Total	Intertidal Upstream		1 15	0	0	252 28	32.25 10.75	21.73 6.83	0 0	C) 0) 0	0 0	.00 .00	.00 .00	42 14
815	Constantine Creek	3 23 93	2.13 2.44 2.74 3.05 3.35 Upstream Upstream Upstream	20 23 30 33 40 80 90 100 120	3 71 0 22 147 48 128 55 16			204 685 522 127 1948 228 0 0	78.32 262.99 200.41 48.76 747.89 87.54 .00 .00	77.91 155.08 135.85 27.44 309.50 55.55 _00 _00 _00	0 0 0 21 0 0 0	41 6 6 6 6 6 6 6 6 6 6		0 0 6 0 160 0 0	.00 .00 2.30 .00 61.43 .00 .00	.00 .00 2.30 .00 61.02 .00 .00	14 14 14 14 14 14 14 14 14
		Tota Tota	l Intertidal l Upstream		243 247	s c 7 c) 4) 1	3486 228	267.68 21.88	79.95 14.44	0 21	41 (6 160	.46 15.36	.46 15.26	70 56

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							Pink	Salmor	1				Chu	m Salmo			
<u> </u>	61				Eg	gs	F	ry	Live	e Fry/m ²	Eg	Igs	Fi	ry	Live	Fry/m ²	-
#	Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	No. of Samples
828	Cook Creek	3 16 93	2.13 2.74 3.35 Upstream	20 30 40 60	26 610 41 341	0 0 28	0 1 1 0	61 499 8 0	23.42 191.58 3.07 .00	21.39 89.17 1.66 .00	0 0 0 0	0 0 0 0	0 0 0 0	0 0 1	.00 .00 .38	.00 .00 .38	14 14 14
		Tota Tota	al Intertidal al Upstream		677 341	0 28	2 0	568 0	72.69 .00	32.61 .00	0 0	0	0	1 0	. 13 . 00	.13 .00	42 14
850	Canoe Creek	3 24 93	2.13 2.74 3.35 Upstream	20 30 40 60	23 19 123 16	0 0 0 0	0 0 1 0	350 136 270 805	134.38 52.21 103.66 309.06	91.69 47.38 63.10 165.89	0 0 0	0 0 0 0	0 0 0 0	0 0 0 1	.00 .00 .00 .38	.00 .00 .00	14 14 14 14
		Tota Tota	l Intertidal il Upstream		165 16	0 0	1 0	756 805	96.75 309.06	39.68 165.89	0 0	0 0	0 0	0 1	.00	.00	42 14
861	Bernard Creek	3 15 93	2.13 2.74 3.35 Upstream	20 30 40 60	14 17 185 125	0 0 0	0 2 4 0	346 447 1576 8	132.84 171.62 605.07 3.07	122.29 96.65 357.20 3.07	0 0 0	0000	0000	0000	.00 .00 .00	.00 .00 .00	14 14 14 14
		Tota Tota	l Intertidal l Upstream		216 125	0 0	6 0	2369 8	303.18 3.07	131.04 3.07	0 0	0 0	0 0	0 0	.00 .00	.00 .00	42 14
Prince	William Sound Su	ummary Tot Tot	al Intertidal al Upstream		18139 11438	18 57	566 217	81274 22234	220.30 160.84	12.51 21.70	215 30	41 0	183 1	3958 1676	10 .73 12.12	3.08 5.54	1983 743

	5		<i></i>			
Order of Collection	Day of Collection	Treatment"	Stream Number	Mean	Std. Error	n
1	1	1	621	0.795	0.0046	24
2	1	2	628	0.835	0.0056	24
3	1	2	637	0.849	0.0285	24
4	1	1	656	0.802	0.0362	24
5	2	2	682	0.807	0.0086	24
6	2	1	695	0.899	0.0058	24
7	3	2	692	0.778	0.0051	18
8	3	1	604	0.889	0.0056	18
9	3	2	663	0.676	0.0095	18
10	3	1	632	0.960	0.0038	18
11	3	2	665	0.747	0.0130	12
12	3	1	653	0.836	0.0065	12
13	4	2	618	0.833	0.0109	24
14	4	1	623	0.918	0.0048	24
15	4	2	618	0.779	0.0226	24
16	4	1	673	0.944	0.0050	24

Appendix B. Estimated mean survival and corresponding standard errors for pink salmon embryos incubated at the Armin F. Koernig hatchery in 1993.

^a Treatment; 1 = control, 2 = oil contaminated