Exxon Valdez Oil Spill Restoration Project Annual Report

Injury to Pink Salmon Embryos in Prince William Sound - Field monitoring

Restoration Project 96191A-1 Annual Report

This annual report has been prepared for peer review as part of the *Exxon Valdez* Oil Spill Trustee Council restoration program for the purpose of assessing project progress. Peer review comments have not been addressed in this annual report.

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Study History: This study originated in March 1989 and continued through February 1991 as Natural Resource Damage Assessment Fish/Shellfish Study Number 2. The project consisted of embryo sampling in the fall and preemergent fry sampling in the spring at oil-contaminated and unimpacted reference streams to determine if the *Exxon Valdez* oil spill affected incubating pink salmon. This work continued in 1992 as Restoration Study Number 60C. At that time the project was expanded to include the previously described field sampling as well as (1) laboratory evaluation of field results through the controlled incubation of pink salmon embryos on oiled substrate (NOAA); (2) an experiment designed to determine if the results observed in the field were due to environmental factors (ADF&G); and (3) a search for evidence of genetic damage (ADF&G). This work was continued as Restoration Projects 93003, 94191, 95191, and this project, Restoration Project 96191. Reports have been finalized for Fish/Shellfish Study Number, Restoration Study Number 60C, and Restoration Projects 93003, 94191 and 95191.

<u>Abstract</u>: We examined pink salmon embryo mortality in intertidal and upstream areas of both oil-contaminated and reference streams in Prince William Sound. Embryo mortality was elevated in oil-affected streams during the falls of 1989, 1990, 1991, 1992 and 1993 (P < 0.028 for all years). However; no statistical difference was observed in the fall of 1994 or 1995 (P > 0.489). We also tested the hypothesis that differences in embryo mortality observed in the field were due to naturally occurring environmental variables that differed systematically between the oil-contaminated and reference streams. Gametes were collected from adults in spawning condition from eight oil-contaminated and eight reference streams, and matings were conducted at a hatchery. The resulting embryos were incubated in controlled environmental conditions. Embryos originating from oil-contaminated streams showed elevated mortalities when compared to the embryos from reference streams in 1993 (P=0.012), but not in 1994 (P=0.343). Results from the controlled incubation study support the results from the field study.

<u>Key Words</u>: Crude oil, embryo mortality, embryos, *Exxon Valdez* oil spill, flow cytometry, genetic damage, *Oncorhynchus gorbuscha*, pink salmon, preemergent fry, Prince William Sound.

<u>Citation</u>:

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EXECUTIVE SUMMARY

This study was designed to monitor recovery of pink salmon Oncorhynchus gorbuscha populations in Prince William Sound that were impacted by the Exxon Valdez oil spill. Embryo mortality and embryo to preemergent fry survival was examined in intertidal and upstream areas of oil-contaminated and unaffected (reference) streams since the spring of 1989. This report focuses upon work performed between October 1, 1995, and September 30, 1996.

Embryo mortality was significantly greater in oiled streams in 1989 and 1990 with the differences observed in all intertidal areas in 1989 and in the highest intertidal area in 1990. These results were consistent with observations of intertidal oiling from other studies. Among oiled streams, all intertidal areas were contaminated in 1989 whereas in 1990 oil remained largely in the upper intertidal zone.

The 1991 evaluation demonstrated a significant difference in embryo mortality between oilcontaminated and reference streams in both the intertidal and upstream zones. This finding was unexpected, as the presence of oil was dramatically reduced in all areas. This result led investigators to hypothesize that:

- 1. oil-induced damage to the 1989 and 1990 broods included deleterious mutations in the germline, or
- 2. incubating embryos continued to be damaged in a physiological manner by an oiled environment even after visually observable oil was gone and this impact was expressed as functional sterility, or
- 3. the observed differences in embryo mortality were due to naturally occurring environmental factors that differed between oiled and reference streams.

All three hypotheses were supportable. Both the genetic-damage and physiological-damage hypothesis seemed credible because past studies confirm that pink salmon embryos take up polycyclic aromatic hydrocarbons (PAHs, Moles et al. 1987), a major component of crude oil, and that these compounds are capable of inducing chromosomal lesions (McBee and Bickham 1988) and influencing endocrine function (Thomas and Budiantara 1995). Pink salmon have an obligate two-year life cycle which results in two genetically isolated lineages, one produced during odd years and the other during even years (Heard 1991). Therefore, genetic or physiological damage induced in one brood year would be expressed in that lineage two years later. The environmental-difference hypothesis seemed credible because environmental factors (wind and currents) determined the distribution of the oil, and those factors might also influence the survivability of salmon embryos incubating intertidally.

Significant differences in embryo mortality were again observed in 1992 and 1993. No statistical difference was detected in 1994 or 1995.

We tested the hypothesis that differences in pink salmon embryo mortality were due to naturally occurring environmental differences. Gametes were collected from adults in spawning condition as they amassed on or near the spawning grounds from eight oilcontaminated and eight reference streams during the 1993 spawning season. The gametes were flown to the Armin F. Koernig hatchery in southwest Prince William Sound where intrastream crosses were made. The resulting embryos from each stream were placed in a common incubator. The pink salmon embryos from oil-contaminated streams showed elevated mortalities when compared to the embryos from reference streams. This finding clearly indicated that the elevated embryo mortalities observed in the field monitoring portion of the study were not due to systematic differences between the incubating environments of oiled and reference streams. This embryo incubation experiment was repeated in 1994, but no significant difference in embryo mortality between oil-contaminated and reference streams was detected. This result is consistent with results obtained from the field monitoring portion of the project in 1994. This study was attempted in 1995 but was incomplete due to lack of spawners in some study streams.

Lack of a statistical difference in pink salmon embryo mortality in 1994 and 1995 between oiled and reference streams demonstrates a possible recovery of the populations that have been monitored since 1989.

INTRODUCTION

Wild salmon play a major role in the Prince William Sound (PWS) ecosystem while also contributing to the region's commercial fisheries. Migrating salmon fry are an important food source in the spring for various mammals, birds, and fishes. Marine mammals prey on the ocean life stages of Pacific salmon while terrestrial mammals and birds, such as bears, river otters, eagles, and gulls depend on salmon for a large portion of their summer diet. Salmon also provide a pathway for transferring nutrients from marine ecosystems to near-shore and terrestrial ecosystems. In recent years, commercial catches of wild salmon have ranged from 10 to 15 million pink salmon and from 0.8 to 1.5 million chum salmon.

Up to 75% of spawning pink (Oncorhynchus gorbuscha) and chum (O. keta) salmon in PWS use intertidal areas (Helle et al. 1964). These areas are highly susceptible to contamination from marine oil spills. Moles et al. (1987) and Rice et al. (1975) found that pink salmon embryos and preemergent fry were adversely affected by exposure to crude oil and that the affect was most acute in intertidal environments. The March 24, 1989 oil spill from the TV Exxon Valdez contaminated many intertidal spawning areas in central and southwest PWS just prior to the spring migration of salmon fry.

Embryo mortality was significantly greater in all intertidal areas of oiled streams in 1989 (P=0.004) and in the highest intertidal area of oiled streams in 1990 (P=0.023, Figure 1) (Sharr et al. 1994a, Bue et al. 1996). These results were consistent with observations of intertidal oil-contamination (Wolfe et al. 1996). Among oiled streams, all intertidal areas were contaminated in 1989 whereas in 1990 oil remained only in the upper intertidal zone.

The 1991 evaluation demonstrated a very significant difference in embryo mortality (P=0.003) between oil-contaminated and reference streams (Figure 1; Sharr et al. 1994a). This finding was unexpected and raised several questions about the source of the elevated mortality in oiled streams, including the possibility that oil-induced damage was transmitted genetically. Petrochemicals have been shown to damage chromosomes (Longwell 1977; McBee and Bickham 1988; Hose et al. *in press*). The pink salmon which spawned during the fall of 1991 were from the 1989 brood year. These fish incubated in oiled gravels during the fall of 1989 and spring of 1990. A pattern of embryo mortality similar to but not as extreme as 1991 was observed in 1992 (P=0.010) and 1993 (P=0.010) (Figure 1; Sharr et al. 1994b and Sharr et al. 1994c). Field sampling in 1994 however, detected no statistical difference between oiled and reference streams (P=0.675, Figure 1; Craig et al. 1996).

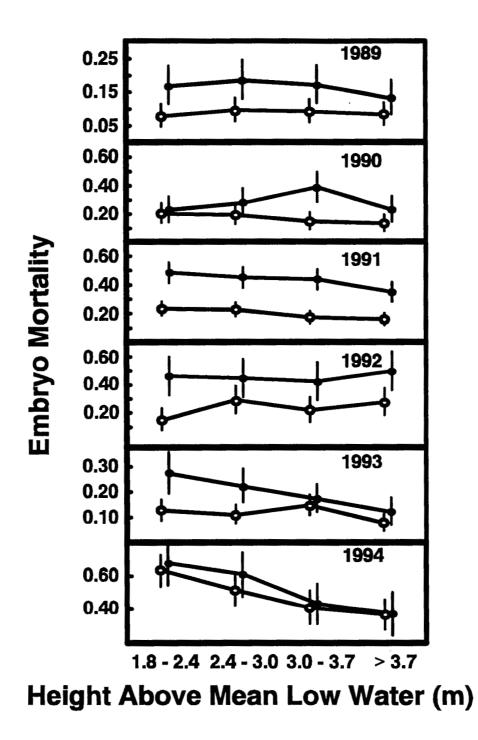


Figure 1. Pink salmon embryo mortality observed during past field sampling. Solid circles indicate the means for oil-contaminated streams (n=10) while open circles identify reference streams (n=15)(90% confidence bounds). Note: Y-axes differ among years.

Three hypotheses have been proposed to explain the differences in embryo mortality observed between oil-contaminated and reference streams:

- 1. oil-induced damage to the 1989 and 1990 broods included deleterious mutations in the germline, or
- 2. incubating embryos continued to be damaged in a physiological manner by an oiled environment even after visually observable oil was gone and this impact was expressed as functional sterility, or
- 3. the observed differences in embryo mortality were due to naturally occurring environmental factors that differed between oiled and reference streams.

All three hypotheses were supportable. Both the genetic-damage and physiological-damage hypothesis seemed credible because past studies confirm that pink salmon embryos take up polycyclic aromatic hydrocarbons (PAHs, Moles et al. 1987), a major component of crude oil, and that these compounds are capable of inducing chromosomal lesions (McBee and Bickham 1988, Biggs et al 1991, Longwell 1977) and influencing endocrine function (Thomas and Budiantara 1995). Pink salmon have an obligate two-year life cycle which results in two genetically isolated lineages, one produced during odd years and the other during even years (Heard 1991). Therefore, genetic or physiological damage induced in one brood year would be expressed in that lineage two years later. The environmental-difference hypothesis seemed credible because environmental factors (wind and currents) determined the distribution of the oil, and those factors might also influence the survivability of salmon embryos incubating intertidally. This embryo mortality study was based on observational data, and as such, we were unable to randomize stream oiling to account for environmental differences between streams. We attempted to address this concern in our original experimental design by selecting reference streams in close proximity to oil-contaminated streams; however, there is a definite oiling pattern in southwest PWS where streams on points which faced northeastward were heavily oiled. Likewise, streams which faced west and southwest were most likely not oiled.

We tested the hypothesis that differences in pink salmon embryo mortality were due to naturally occurring environmental differences. Gametes were collected from adults in spawning condition as they amassed on or near the spawning grounds from eight oilcontaminated and eight reference streams during the 1993 spawning season (Sharr et al. 1994c). The gametes were flown to the Armin F. Koernig hatchery in southwest Prince William Sound where intrastream crosses were made. The resulting embryos from each stream were placed in a common incubator. The pink salmon embryos from oil-contaminated streams showed elevated mortalities when compared to the embryos from reference streams (P=0.012). This finding clearly indicated that the elevated embryo mortalities observed in the field monitoring portion of the study were not due to systematic differences between the incubating environments of oiled and reference streams. This embryo incubation experiment was repeated in 1994, but no significant difference in embryo mortality between oilcontaminated and reference streams was detected (P=0.343; Craig et al. 1996). This result is consistent with results obtained from the field monitoring portion of the project in 1994. No difference in embryo to preemergent fry survival between oil-contaminated and reference streams has been observed since the initiation of the study in 1989 (Figure 2; Sharr et al. 1994a, 1994b, and 1994c). We expected embryo to preemergent fry survival to be reduced in oiled streams given that an increase in embryo mortality had been detected. This result can potentially be explained by (1) compensation in the environment, or (2) problems in the experimental design. Geiger et al. (1996) found no evidence to suggest that compensation in the intragravel life stages is playing a role in determining the number of emerging fry for the years in the study. We believe that the experimental design is inadequate for detecting differences in embryo to preemergent fry survival. The power analysis for the survival from embryo to preemergent fry test indicated statistical power was adequate to detect a biologically meaningful difference if present. However, unexpected changes in stream characteristics may have prevented sampling the same areas for embryos in the fall and fry in the spring. Some intertidal stream segments were found to migrate along the beach, especially if the beach is usually exposed to winter storms. The magnitude of these changes was unexpected when this study was designed and initiated.

This project continued to monitor salmon streams for embryo mortality as well as evaluate the environmental-difference hypothesis.

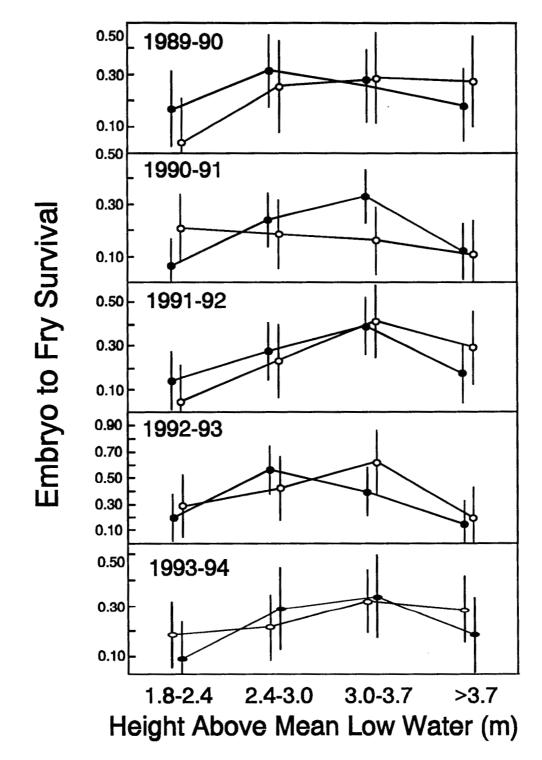


Figure 2. Pink salmon embryo to preemergent fry survival observed during past field sampling. Solid circles indicate the means for oil-contaminated streams (n=10) while open circles identify reference streams (n=15)(90% Confidence bounds). Note: Y-axes differ among years.

OBJECTIVES

Recovery Monitoring of Injury to Pink Salmon Embryos in Prince William Sound

1. Test for differences in mortality of pink salmon embryos between oilcontaminated and reference streams.

Verification of Injury to Pink Salmon Gametes in Prince William Sound

1. Determine if the increased pink salmon embryo mortalities observed in oiled streams can be attributed to systematic environmental differences between the oil-contaminated and reference streams.

METHODS

Recovery Monitoring of Injury to Pink Salmon Embryos in Prince William Sound

Study Sites

This project concentrated on populations inhabiting the southwestern portions of PWS; although, streams from Montague Island and eastern PWS were sampled to provide a sound-wide perspective (Figure 3).

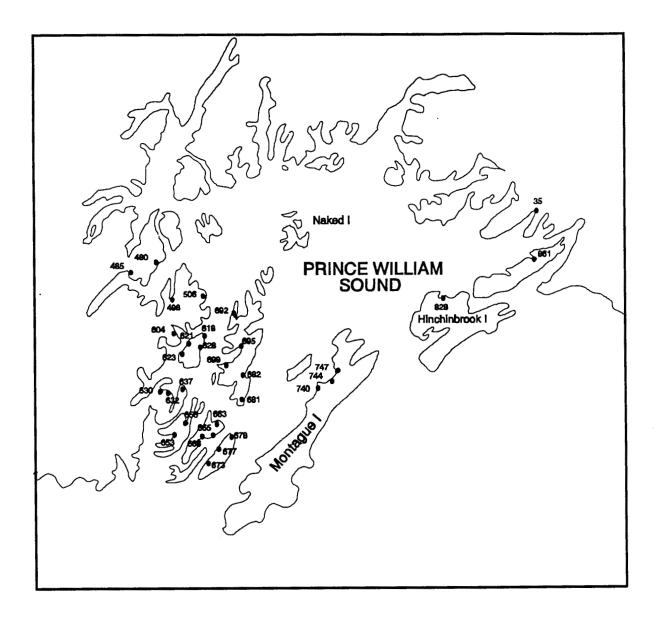


Figure 3. Streams examined during the 1989, 1990, 1991, 1992, 1993, 1994, and 1995 pink salmon preemergent fry and egg deposition surveys.

These streams were selected for the following reasons:

- 1. They have significant spawning populations in both odd and even years.
- 2. They are accessible for sampling in most years.
- 3. They are representative of oiled or reference sites in the oil-impacted area.

Sample Design

The methods used for embryo sampling are described by Craig et al. (1995) and Sharr et al. (1994a and 1994b). Sampling was stratified by tide zone to control for possible differences in salinity, temperature, predation, or a combination of these factors. Zone boundaries were established with a surveyor's level and stadia rod and staked prior to sampling. Four zones were sampled in each stream whenever possible: 1.8 - 2.4 m, 2.4 - 3.0 m, 3.0 - 3.7 m above mean low water, and upstream of mean high tide (3.7 m). No sampling was done below the 1.8 - 2.4 m zone because mortality was expected to be high (Helle et al. 1964).

Separate linear transects were established for each zone on the embryo surveys. Although most transects were 30.5 m long, some were shorter due to steep stream gradients. Transects were placed in riffle areas where spawning was observed during escapement surveys conducted for F/S 1. Transects ran diagonally across the river starting downstream against the left bank and moved upstream to the right bank. A map drawn for each stream indicated the tide zones and transect locations in relation to major landmarks. Each embryo transect was photographed and marked with surveyor's flagging to insure that future transects could be located in the same area of the stream.

Fourteen circular samples (0.186 m²) were systematically collected along each transect. The number of digs was a compromise between reducing variance and the practicality of conducting the study. Fewer digs were completed on narrow stream channels to avoid excessive sampling of the stream. Streams that split into two or more channels within a zone were sampled either by allocating digs among channels based on spawner distribution observed during F/S 1 or, where spawner distribution was unknown, by an equal allocation.

The following data were collected for each tide zone transect during both embryo and fry sampling:

- 1. Sample date.
- 2. Sample tide zone.
- 3. Start and stop time for the tide zone transect.
- 4. Numbers of live and dead fry and embryos for each species in each dig.

Pink salmon embryos were separated from chum O. keta and coho O. kisutch salmon embryos by their smaller size. Chum salmon embryos were separated from coho salmon embryos by

their greater development and different coloration. An embryo was considered dead if it was opaque or discolored with coagulated lipids. Fry were considered dead only if decomposition was evident, because sampling often killed fry.

Data Analysis

Numbers of live and dead embryos and fry were summarized by date, stream, level of hydrocarbon impact, and stream zone. Densities of live embryos for stream i, zone j in m^2 (E_{ii}) were estimated by:

$$\hat{\mathbf{E}}_{ij} = \frac{\Sigma L E_{ijk}}{0.3 n_{ij}} , \qquad (1)$$

where LE_{ijk} is the number of live embryos found in the kth dig, in stream i, zone j, and n_{ij} is the number of digs from stream i, zone j. Densities of dead embryos were calculated using the same estimator with appropriate substitutions.

Pink salmon embryo mortality was estimated for each stream using the following relationship:

$$\hat{M}_{ij} = \frac{\Sigma (DE_{eijk} + DF_{eijk})}{\Sigma (LE_{eijk} + DE_{eijk} + LF_{eijk} + DF_{eijk})} , \qquad (2)$$

where DE_{eijk} , DF_{eijk} , LE_{eijk} , and LF_{eijk} are the number of dead embryos, dead fry, live embryos, and live fry for the kth dig from stream i, zone j, collected during embryo dig e, respectively.

The Logit transform of embryo mortality [ln (odds)], i.e.

$$\text{Logit}_{ij} = \ln \left[\frac{\Sigma (\text{DE}_{eijk} + \text{DF}_{eijk})}{\Sigma (\text{LE}_{eijk} + \text{LF}_{eijk})} \right] , \qquad (3)$$

was used.

Differences in embryo mortality were examined using a mixed effects two-factor experiment with repeated measures on one factor (Neter et al. 1990):

$$Y_{ijk} = \mu_{...} + O_i + Z_j + (OZ)_{ij} + S_{k(i)} + e_{(ijk)}.$$
 (4)

The two treatments were level of oiling, $(O_i, 2 \text{ levels}; \text{ oiled and reference})$, and height in the intertidal zone (Z_j , 4 levels; 2.1, 2.7, and 3.4 m above mean low water, and upstream) both fixed effects. The data were blocked by stream ($S_{k(i)}$), a random effect nested within level of oiling. The interaction of level of oiling and height in the intertidal zone was also examined.

Equality of variances was tested using the F_{max} -test (Sokal and Rohlf, 1969), while normality of error terms was visually assessed using normal quantile-quantile and box plots (Chambers et al. 1983). Arcsin square root, logit, log, and square root transforms were examined if the data indicated non-constant variances or non-normal error terms. Tests of homogeneity of between treatment covariance matrices and the degree of sphericity of the pooled covariance matrix were effected. Four contrasts (oil vs. reference for the four stream zones) and corresponding Bonferroni family confidence intervals ($\alpha = 0.10$ overall) were estimated if a significant difference due to oiling was detected. The SAS (SAS Institute Inc. 1988) General Linear Models Procedure was used to analyze the data.

Stream oiling was assessed through visual observations of the stream and the adjacent area. The observations were supported by photographs, observation maps, and hydrocarbon analysis of mussels (*Mytilus* sp.) collected near stream mouths. These data were collected as part of another Natural Resource Damage Assessment study (Sharr et al. 1994a).

Verification of Injury to Pink Salmon Gametes in Prince William Sound

Embryo Rearing

The environmental difference/genetic damage hypotheses were evaluated through an experiment in which embryos from oil-contaminated and reference streams were incubated in a common environment. Gametes from 30 male and 30 female pink salmon were collected from each of eight oil-contaminated and eight reference streams in southwestern PWS (Figure 4). Each oil-contaminated stream was paired with a reference stream based on similarity of geographic location and physical characteristics. Paired streams were sampled on the same day and gametes flown to the Armin F. Koernig (AFK) hatchery.

Gamete collection techniques were identical at each stream. Adults were captured at low tide in the stream mouth using a 30-m hand operated beach seine. Only gametes from ripe individuals (adults that readily extruded eggs or sperm when gently massaged) were taken. Eggs from individual females (approx. 1500 per female) were removed by excising the abdominal wall, allowing eggs to flow directly into 1-L zip-lock plastic bags, and packed on cotton towels over a 10-cm layer of wet ice in insulated ice chests. Sperm from individual males (2-3-ml) was placed into 15-ml plastic centrifuge tubes which were then capped and placed on ice in the same chests as the females for that stream. After collection was complete, gametes were flown back to AFK Hatchery (an average 10 minute flight time) while gametes from the next stream were collected.

The construction of stream specific embryo pools consisting of all single-pair crosses (30 x 30 = 900) was begun immediately upon arrival of the gametes at the hatchery. Crosses were made by first placing 5-ml of eggs (approximately 30 eggs) from each female into each of 30, 0.47-L cups (each cup contained a teaspoon of eggs from each female). Each cup was then fertilized by a single male using 100 μ l of sperm followed by 100-ml of freshwater (8 °C) to initiate fertilization. This procedure provided each male an equal opportunity to fertilize eggs from each female. The fertilized eggs were allowed to sit for approximately 3-min after which they were recombined into a 3-L plastic container (maintained at 8 °C) and gently rinsed and mixed with freshwater three times.

The matings from each day were placed into one of four stacks of Heath trays (FAL/Heath Tray, Tacoma, Washington, U.S.A.). Six trays within each of the four stacks were divided into 16 compartments (four rows by four columns) using plastic strips, providing 96 compartments for replicated incubation. Each strip was sealed to the tray to prevent mixing of embryos and larvae between compartments. Twenty four replicates of approximately 580 embryos (100-ml of embryos) each were randomly collected from the stream-specific embryo pools and loaded into separate compartments using a random loading scheme on sampling days.

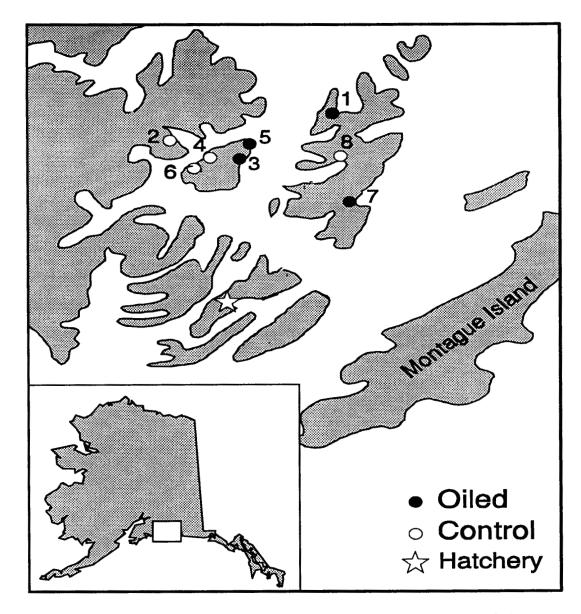


Figure 4. Map of study area in southwestern Prince William Sound, Alaska. Index number next to stream locator indicates the order of gamete collection.

A technician was stationed at the hatchery during the three months of the experiment to perform normal fish culture duties and collect mortality data. The technician was made aware of the day of collection for record keeping but did not know which incubator compartments represented oil-contaminated or reference streams.

Dead embryos in each compartment were counted and removed 36 hours after fertilization, after which trays were undisturbed for four weeks. Water flow to each of the four incubator stacks was maintained at four gpm. Each incubator stack received a 20-ppt sodium chloride bath for 20 minutes duration twice per week to control fungus infestations on the embryos. Water temperatures during incubation ranged from 5 ° to 11 °C.

Mortality of embryos at the eyed stage (the point at which a distinct embryo eye could be seen through the chorion) was recorded at 350 temperature units (T.U.; 1 T.U. = 1 °C above 0 °C for a 24-hr period). Embryos at this stage were siphoned out of their compartments using Tygon tubing (10 mm inside diameter) and allowed to drop 10-12 cm into a container of freshwater. The resulting physical shock caused coagulation of yolk material in undeveloped embryos, allowing easier identification and removal. Live and dead embryos were gently placed back into their original compartments after siphoning. The live embryos were counted, and the dead embryos were removed and counted.

Mortality was again recorded after the embryos had completely hatched (770 T.U.). In addition, the number of abnormal larvae (deformities of the head, body, or tail) in each compartment was recorded. All larvae were destroyed after hatching.

The statistical difference in mortality (Y) due to oil contamination (O) was evaluated using a blocked (day; D) analysis of variance, i.e.

$$Y_{ijk} = \mu_{..} + D_i + O_j + \varepsilon_{ijk} \qquad . \tag{5}$$

RESULTS

Recovery Monitoring of Injury to Pink Salmon Embryos in Prince William Sound

Thirty-one streams were sampled between October 2 and October 25, 1995 for embryos. Mean embryo densities for the 1995 egg deposition survey were 340.15 eggs per m^2 in the intertidal zones and 578.66 eggs per m^2 in the upstream (Appendix A). The 1995 embryo mortality data indicated no significant difference between the oil-contaminated and reference streams (P=0.489; Figure 5). No significant zone effect (P=0.280) or oil-by-zone interaction was found (P=0.318). The overall mean embryo mortalities for the oil contaminated and reference streams were 0.283 and 0.234.

Verification of Injury to Pink Salmon Gametes in Prince William Sound

Gamete collection began with four streams on August 28, 1995. Four streams were also sampled the next day, August 29, 1995. Pink salmon runs failed to materialize in the remaining streams and the collections were discontinued.

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Embryo mortalities from oil-contaminated and reference streams were similar when both groups were incubated in the controlled environment of the AFK Hatchery (Figure 6; Appendix B). The Difference in mortality between reference and oil-contaminated streams was not tested due to the lack of statistical power.

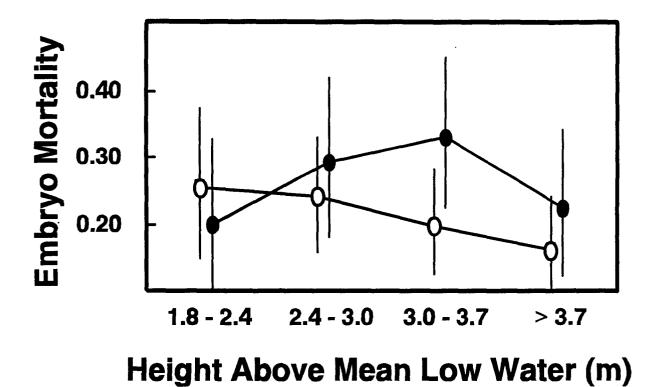


Figure 5. Mean pink salmon embryo mortality and corresponding 90% confidence bounds by tide zone for oil-contaminated and reference streams in Prince William Sound, 1995. Solid circles represent data from ten oil-contaminated streams, and open circles represent data from 15 reference streams.

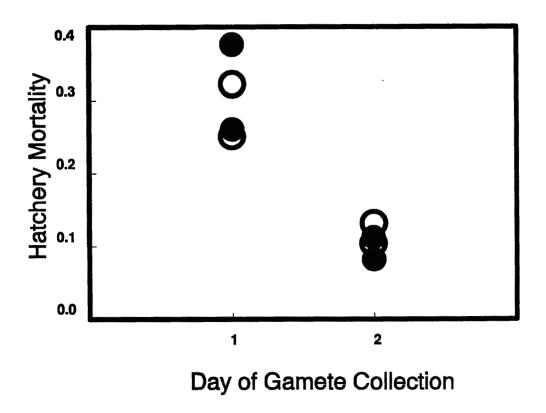


Figure 6. Mean pink salmon embryo mortality observed under hatchery conditions in 1995. Solid circles indicate oil-contaminated streams while open circles indicate reference streams.

DISCUSSION

Pink salmon embryos that incubated in oil-contaminated spawning areas in PWS appeared to have been adversely affected by the *Exxon Valdez* oil spill up until 1994. Sharr et al. (1994a, 1994b and 1994c) found increased pink salmon embryo mortalities from 1989 through 1993 (Figure 1). However; embryo mortality was not significantly different between oil-contaminated and reference streams in 1994 (Figure 1) and 1995 (Figure 5). Lack of a significant difference in 1994 and 1995 between oiled and reference streams demonstrates a possible recovery of the populations that have been monitored since 1989.

The pink salmon that spawned during the fall of 1991 incubated in oil-contaminated streams during the winter of 1989-1990, the first winter after the spill. Likewise, pink salmon that spawned during the fall of 1992 incubated in oiled stream gravel during the winter of 1990-1991. Sharr et al. (1994a) found significantly elevated embryo mortalities in oil-contaminated streams during the fall of 1989 and 1990, and the surviving embryos may have sustained sublethal damages which could have been manifested in the form of functional sterility in 1991, 1992 and 1993. Polycyclic aromatic hydrocarbons have been shown to influence endocrine function (Thomas and Budiantara 1995) as well as damage chromosomes (Longwell 1977, McBee and Bickham 1988, Hose et al. *in press*, Biggs et al 1991), although documentation of germline damage is yet to be reported. In 1994 the returning adults were two generations away from the 1990 brood and showed no statistical difference in mortality in the field or the controlled incubation experiment. In 1995 the returning adults were three generations away from the 1989 brood and again showed no statistical difference in the field study. Mortality differences in oiled and reference streams seem to be returning to normal levels, lending credit to a post-spill damage hypothesis with later recovery.

Environmental differences between oil-contaminated and reference streams does not seem to be a confounding factor because results from the controlled incubation experiment in 1993 and 1994 mirrored the results of the field study. The data for the 1995 controlled incubation experiment were not analyzed because of a lack of statistical power; although, the oiled and reference stream mortalities again look similar (Figure 6). The gametes used in these studies were never in direct contact with a stream; although, the adults which produced them had incubated in the natal streams.

These data do not prove that the observed differences in embryo mortality between oilcontaminated and reference streams were caused by exposure to hydrocarbons. It is possible that the streams which were oiled historically had lower embryo survival. However, this seems unlikely because mortality between oil-contaminated and reference streams was not significantly different in 1994 or 1995. The controlled oiling experiment conducted by the National Marine Fisheries Service (Project 95320C) will provide laboratory evidence to further clarify interpretation of these field data.

CONCLUSIONS

Embryo mortalities were elevated in oil contaminated streams from 1989-1993, but no statistical difference between oil-contaminated and reference streams was observed in 1994 and 1995.

Mortalities in an incubation experiment were elevated for embryos from oiled streams in 1993, but no statistical difference between embryos from oil-contaminated and reference streams was detected in 1994.

ACKNOWLEDGEMENTS

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Appendix A. Summary of pink and chum salmon egg dig data from Prince William Sound, 1995.

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							Pink	Salmon					Chur	Salmon			_
					Emb	ryos	Live	Embryos/n	n² Fr	У	Embry	05	Live Emb	ryos∕m²	Fr	1	_
Stream #	Stream Name	Dat	Height in e Tidal Zone(m)	Loc	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
35	Koppen Creek	10 2	95 9.0 11.0 Upstream	30 40 60	6003 9730 7816	418 792 670	160.48 304.07 257.23	60.60 96.83 71.14	0 0 0	0 0 0	757 31 29	112 93 3	43.00 35.71 1.15	27.50 32.13 .61	0 0 0	0 0 0	14 14 14
			Cotal Intertidal Cotal Upstream		15733 7816	1210 670	232.28 257.23	57.73 71.14	0 0	0 0	788 29	205 3	39.35 1.15	20.76 .61	0 0	0 0	28 14
480	Mink Creek	10 9	95 7.0 9.0 11.0 Upstream	20 30 40 60	5 127 50 6	1 1175 371 3	.38 451.12 142.44 1.15	.38 249.49 139.56 1.15	0 0 0 0	0 0 0 0	0 3 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0 0	0 6 1 0	14 14 14 14
			Cotal Intertidal Cotal Upstream		182 6	1547 3	197.98 1.15	97.47 1.15	0 0	0	3 0	0 0	.00	.00 .00	0 0	7 0	42 14
484	E. Finger Creek	10 16	95 7.0 9.0 11.0	20 30 40	7 317 323	183 1889 80	70.26 725.24 30.71	38.93 347.85 18.45	0 0 0	0 0 0	0 0 6	0 0 0	.00 .00 .00	.00 .00 .00	0 0 0	0 0 3	14 14 14
			'otal Intertidal 'otal Upstream		647 0	2152 0	275.40 .00	124.33 .00	0 0	0 0	6 0	0 0	.00 .00	.00 .00	0 0	3 0	42 0

Appendix A.	Summary of	of pink and	d chum s	almon eg	g dig	data from	Prince	William	Sound,	1995.

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								Pink	Salmon					Chum	Salmon			
						Emb	ryos	Live 1	Embryos/m	² Fr	Y	Embry	os	Live Emb	ryos/m²	Fry	ť	_
Stream #	Stream Name	Dat		Height in Fidal Zone(m)	Loc	Dead	Live	e Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Sample
185	W. Finger Creek	10 8	95	7.0	20	5	373	143.21	132.72	0	0	102	91	34.94	24.63	1	138	14
				9.0 11.0 Upstream	30 40 60	135 234 198		44.92 1161.38 1601.37	32.65 335.58 463.30	0 0 0	0 2 1	15 24 1	0 85 0	.00 32.63 .00	.00 31.81 .00	0 0	0 186 0	14 14 14
				Intertidal Upstream		374 198		449.84 1601.37	141.74 463.30	0 0	2 1	141 1	176 0	22.52 .00	13.31 .00	1 0	324 0	42 14
198 1	McClure Creek	10 8	95	7.0 9.0 11.0 Upstream	20 30 40 60	221 1190 762 284		630.03 1360.26 1758.78 278.35	280.04 377.83 445.86 138.80	0 0 0	0 0 2 0	23 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
				Intertidal Upstream		2173 284	9765 725	1249.69 278.35	222.97 138.80	0 0	2 0	23 0	0 0	.00	.00 .00	0 0	0 0	42 14
06 L	Loomis Creek	10 10	95	7.0 9.0 11.0 Upstream	20 30 40 60	188 1500 3364 622	896 1866 1187 768	344.00 716.41 455.72 294.86	170.25 164.01 126.54 142.29	0 0 0 0	0 0 0	0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00	0 0 0	0 0 0	14 14 14 14
				Intertidal Upstream		5052 622	3949 768	505.38 294.86	90.51 142.29	0 0	0 0	0 0	0 0	.00 .00	.00	0 0	0	42 14

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							Pink	Salmon					Chum	Salmon			_
					Emb	ryos	Live	Embryos/n	n² Fr	Y	Embry	os	Live Embr	yos∕m³	Fry	ł	
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
604	Erb Creek	10 18 95	7.0 7.0 9.0 11.0 Upstream	20 23 30 40 60	8 10 1 294 5	1 27 3 887 38	.77 20.73 1.15 340.54 14.59	.77 18.96 .61 226.96 12.26	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	.00. .00 .00	.00 .00 .00 .00	0 0 0 0	0 0 0 0	7 7 14 14 14
			Intertidal Upstream		313 5	918 38	117.48 14.59	77.85 12.26	0 0	0 0	0 0	0 0		.00 .00	0 0	0 0	42 14
618 .	Junction Creek	10 11 95	7.0 9.0 11.0 Upstream	20 30 40 60	3 5 0 7	1 0 0 24	.45 .00 .00 10.75	.45 .00 .00 10.27	0 0 0 0	0 0 0	0 0 0 0	0 0 0 0	.00 .00	.00 .00 .00 .00	0 0 0 0	0 0 0 0	12 12 12 12
			Intertidal Upstream		8 7	1 24	.15 10.75	.15 10.27	0 0	0 0	0 0	0 0		.00	0 0	0 0	36 12
621	Totemoff Creek	10 17 95	7.0 9.0 11.0 Upstream	20 30 40 60	908 404 124 1092	148 894 179 2015	56.82 343.23 68.72 773.62	24.04 160.88 48.80 212.51	0 0 0	3 4 4 180	0 0 0 0	0 0 0 0	.00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		1436 1092	1221 2015	156.26 773.62	58.95 212.51	0 0	11 180	0 0	0 0		.00 .00	0 0	0 0	42 14

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							Pink	Salmon					Chum	Salmon			
					Emb	oryos	Live H	Embryos/m	² Fr	У	Embryo	55	Live Embr	yos/m²	Fry	Y	_
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	e Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
623	Brizgaloff Creek	10 17 95	7.0 9.0 11.0 Upstream	20 30 40 60	393 179 114 2178	442 704 356 1217	169.70 270.29 136.68 467.24	99.84 175.58 86.69 161.22	0 0 0 0	6 2 0 3	0 3 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		686 2178	1502 1217	192.22 467.24	72.00 161.22	0 0	8 3	3 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
628	Chenega Creek	10 10 95	7.0 9.0 11.0 Upstream	20 30 40 60	16 93 350 364	15 246 3271 1397	5.76 94.45 1255.83 536.35	2.22 30.04 260.63 172.67	0 0 0	0 0 0 0	0 0 0 4	0 0 8	.00 .00 .00 3.07	.00 .00 .00 3.07	0 0 0	0 0 0 1	14 14 14 14
			Intertidal Upstream		459 364	3532 1397	452.01 536.35	123.23 172.67	0 D	0 0	0 4	0 8	.00 3.07	.00 3.07	0 0	0 1	42 14
630	Bainbridge Creek	10 13 95	7.0 9.0 11.0 Upstream	20 30 40 60	3 247 581 556	2 457 1807 3188	.77 175.46 693.76 1223.96	.52 166.48 239.52 316.35	0 0 0	0 0 213 8	0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0 0	14 14 14 14
			Intertidal Upstream		831 556	2266 3188	289.99 1223.96	105.38 316.35	0 0	213 8	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
632	Claw Creek	10 12 95	7.0 9.0 11.0 Upstream	20 30 40 60	6 22 493 3	6 829 3823 221	2.30 318.28 1467.76 169.70	.93 313.32 408.08 111.06	0 0 0	0 2 1 0	0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 7
			Intertidal Upstream		521 3	4658 221	596.11 169.70	194.03 111.06	0 0	3 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 7
637	Pt. Countess	10 12 95	7.0 9.0 11.0 11.0 Upstream Upstream	20 30 41 42 61 62	1643 672 228 156 282 41	567 282 816 93 163 265	217.69 108.27 626.57 71.41 125.16 203.48	115.93 47.74 475.19 39.27 51.44 136.36	0000000	0 0 0 0 0	0 1 0 0 0	0 1 0 0 0	.00 .38 .00 .00 .00	.00 .38 .00 .00 .00		0 0 0 0 0	14 14 7 7 7 7
			Intertidal Upstream		2699 323	1758 428	224.98 164.32	89.83 70.85	0 0	0 0	1 0	1 0	.13 .00	.13 .00	0 0	0 0	42 14

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							Pink	Salmon					Chum	Salmon			
					Emb	ryos	Live	Embryos/1	n² Fr	Y	Embry	os	Live Embr	yos/m²	Fr	1	-
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
653	Vogg Crock	10 15 95	7.0	20	5	21	8.06	3.17	0	0	0	0	.00	.00	0	0	14
53	ogg Creek	10 15 95	9.0	31	0	0	.00	.00	0 0 0	0	Ő	0	.00	.00	0	0 0	7
			9.0	32	5	4	3.07	2.30	0	0	0	0	.00	.00	0	0	7
			11.0	40 60	0 71	100	.38	.38	0	0	0	0	.00	.00 .00	0	0	14 14
			Upstream	60	/1	486	186.59	106.95	U	U	U	U	.00	.00	U	0	14
		Tota	l Intertidal		10	26	3.33	1.23	0	0	0	0	.00	.00	0	0	42
			l Upstream		71	486	186.59	106.95	0	0	0	0	.00	.00	0	0	14
656	Halverson Creek	10 14 95	7.0	20	163	1148	440.75	270.07	0	0	0	0	.00	.00	0	0	14
000	nutrerben eren		9.0	30	67	860	330.18	194.18	Ō	Ō	ō	0	.00	.00	õ	0	14 14
			11.0	40	838	2223	853.47	287.46	0	0	0	0	.00	.00	0	0 0	14 14
			Upstream	60	380	2141	821.99	175.71	0	1	0	0	.00	.00	0	0	14
		Tota	l Intertidal		1068	4231	541.47	147.19	0	0	0	0		.00	0	0	42
		Tota	l Upstream		380	2141	821.99	175.71	0	1	0	0	.00	.00	0	0	14

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							Pink	Salmon					Chum	Salmon			_
					Emb	oryos	Live	Embryos/n	1² Fr	Ŷ	Embry	05	Live Embr	yos/m²	Fr	ł	
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	e Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
663	Shelter Bay	10 13 95	7.0 9.0 11.0 Upstream	20 30 40 60	19 0 128 164	317 9 489 3281	141.99 4.03 219.03 1469.61	84.97 1.89 95.48 589.97	0 0 0	0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0 0	0 0 0	12 · 12 12 12 12
			Intertidal Upstream		147 164		121.68 1469.61	44.02 589.97	0 0	0 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	36 12
665	Bjorne Creek	10 19 95	7.0 9.0 11.0 Upstream	20 30 40 60	67 402 316 292	327 453 692 1267	125.54 173.92 265.68 486.44	65.95 64.84 136.82 129.55	0 0 0	0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0 0	14 14 14 14
			Intertidal Upstream		785 292	1472 1267	188.38 486.44	54.45 129.55	0 0	0 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
666	O'Brien Creek	10 15 95	7.0 9.0 11.0 Upstream	20 30 40 60	1 11 29 140	3 52 370 317	1.15 19.96 142.05 121.71	.61 13.82 79.97 67.47	0 0 0 0	0 0 0 0	0 0 0	0 0 0	.00 .00 .00	.00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		41 140	425 317	54.39 121.71	28.13 67.47	0 0	0 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
673	Falls Creek	10 25 95	7.0 9.0 11.0 Upstream	20 30 40 60	181 18 27 136	706 87 291 1472	271.05 33.40 111.72 565.14	183.59 28.81 71.92 184.46	0 0 0	0 0 85 0	0 0 0 0	0 0 0	.00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		226 136	1084 1472	138.73 565.14	65.60 184.46	0 0	85 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
677	Hayden Creek	10 25 95	7.0 7.0 9.0 11.0 11.0 Upstream Upstream	21 22 31 32 41 42 61 62	8 8 17 13 27 0 0 1	103 355 314 51 22 0 7 1	79.09 272.59 241.11 39.16 16.89 .00 5.38 .77	68.41 170.56 162.77 22.70 12.10 .00 5.38 .77	0 0 0 1 0 0	0 1 0 1 0 0 0 0		0 0 0 0 0 0 0	.00 .00 .00 .00 .00 .00 .00	.00 .00 .00 .00 .00 .00 .00	0 0 0 0 0 0 0	0 0 0 0 0 0	7 7 7 7 7 7 7 7

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							Pink	Salmon				•	Chum	Salmon			
					Emb	ryos	Live	Embryos/n	n² Fr	У	Embry	05	Live Embr	yos∕m³	Fr	¥	-
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
			al Intertidal al Upstream		73 1	845 8	108.14 3.07	42.09 2.69	1 0	2 0	0 0	0 0	.00	.00 .00	0 0	0 0	42 14
578 <u>s</u>	Sleepy Bay	10 13 95	7.0 9.0 11.0 Upstream	20 30 40 60	2 35 15 69	38 1354 41 640	17.02 606.48 18.36 286.67	$11.65 \\ 223.08 \\ 16.43 \\ 148.94$	0 0 0	0 0 0 0	0 0 0 0	0 0 0	.00	.00 .00 .00 .00	0 0 0	0 0 0 0	12 12 12 12
			al Intertidal al Upstream		52 69	1433 640	213.95 286.67	86.35 148.94	0 0	0 0	0 0	0 0		.00	0 0	0 0	36 12
681	Hogan Bay	10 21 95	7.0 9.0 11.0 Upstream	20 30 40 60	1248	3581	259.15 1033.15 1374.85 1020.48	163.23 390.31 396.31 327.59	0 0 0	0 0 0	0 0 0	0 0 0 0	.00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			l Intertidal 1 Upstream			6947 2658	889.05 1020.48	202.07 327.59	0 0	0	0 0	0 0	.00 .00	.00	0 0	0 0	42 14

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	Stream Name						Pink	Salmon			Chum Salmon						
					Emb	Embryos		Live Embryos/m		m² Fry		05	Live Embr	yos/m²	Fr	ł	_
Stream #		Date	Height in Tidal Zone(m)	Loc	Dead	Live	e Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
682	Snug Harbor	10 21 95	7.0 9.0 11.0 Upstream	20 30 40 60	1341 1145 1684 4927	2648 6693	1057.34 1016.64 2569.63 3060.68	287.04 192.07 506.75 628.55	0 0 0	0 0 0 3	0 0 0 0	0 0 0 0	.00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		4170 4927		1547.87 3060.68	229.10 628.55	0 0	0 3	0 0	0 0		.00	0	0	42 14
692	Herring Bay	10 9 95	7.0 9.0 11.0 Upstream	20 30 40 60	613 722 1492 162	1138 907 2277 790	436.91 348.22 874.21 303.30	200.16 98.98 177.79 135.50	0 0 0	0 0 0	0 0 0 0	0 0 0 0	.00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		2827 162	4322 790	553.11 303.30	99.50 135.50	0 0	0 0	0 0	0 0		.00 .00	0 0	0 0	42 14
695	Port Audrey	10 11 95	7.0 7.0 9.0 11.0 Upstream	21 22 30 40 60	105 54 876 503 27	183 73 1023 564 517	140.52 56.05 392.76 216.54 198.49	65.64 30.27 136.43 117.01 162.24	0 0 0 0	0 0 0 0	0 0 3 0 0	0 0 0 0	.00 .00	.00 .00 .00 .00	0 0 0 0	0 0 4 0 0	7 7 14 14 14
			Intertidal Upstream		1538 27	1843 517	235.86 198.49	62.56 162.24	0 0	0 0	3 0	0 0	.00	.00	0	4 0	42 14

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							Pink	Salmon				•	Chum	Salmon			_
					Emb	огуов	Live 1	Embryos/m	² Fr	У	Embry	os	Live Embr	yos/m²	Frj	ŗ	
Stream #	Stream Name	Date	Height in Tidal Zone(m)	Loc	Dead	Live	e Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
699	Cathead Bay	10 11 95	7.0 9.0 11.0 Upstream	20 30 40 60	1043 465 429 598	578 464 1186 2859	221.91 178.14 455.34 1097.65	72.10 78.02 169.43 347.59	0 0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00	0 0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		1937 598	2228 2859	285.13 1097.65	67.73 347.59	0 0	0 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
740 1	Kelez Creek	10 22 95	7.0 9.0 11.0 Upstream	20 30 40 60	69 174 276 167	442 596 638 1328	169.70 228.82 244.95 509.86	122.93 93.80 139.51 238.04	0 0 0 0	0 0 0	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		519 167	1676 1328	214.49 509.86	67.89 238.04	0 0	0 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
744	Wilby Creek	10 22 95	7.0 9.0 11.0 Upstream	20 31 40 60	655 111 180 46	16 131 2153 165	6.14 50.29 826.60 63.35	3.13 20.44 240.36 46.69	0 0 0	0 0 0 0	0 0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		946 46	2300 165	294.35 63.35	98.05 46.69	0 0	0 0	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
747	Cabin Creek	10 23 95	7.0 9.0 11.0 Upstream	20 30 40 60	267 1254 1075 839	16 351 1665 2652	6.14 134.76 639.24 1018.18	4.54 100.31 206.30 326.22	0 0 0 0	0 0 38 1	0 0 0 0	0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 0	14 14 14 14
			Intertidal Upstream		2596 839	2032 2652	260.05 1018.18	85.93 326.22	0 0	38 1	0 0	0 0	.00 .00	.00 .00	0 0	0 0	42 14
828	Cook Creek	11 2 95	7.0 9.0 11.0 Upstream	40	8 1190 1022 1124		1.92 260.69 1089.97 1962.64	.91 100.54 313.94 509.25	0 0 0	0 1 192 6	0 0 17 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0	0 0 1 0	14 14 14 14
			Intertidal Upstream		2220 1124	3523 5112	450.86 1962.64	129.38 509.25	0 0	193 6	17 0	0 0	.00	.00	0 0	1 0	42 14

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Appendix A. (page 10 of 10)

					Loc		Pink Salmon						Chum Salmon					
Stream #				Height in Tidal Zone(m)		Embryos		Live Embryos/m		m² Fry		Embryos		Live Embryos/m ²		Fry		
	Stream Name		Date			Dead	Live	Mean	SE	Dead	Live	Dead	Live	Mean	SE	Dead	Live	No. Samples
861	Bernard Creek	10	3 95	7.0 9.0 11.0 Upstream	20 30 40 60	33 4514 2192 1620	2 1108 151 1906	.77 425.39 57.97 731.77	.52 189.25 22.71 254.02	0 0 0 0	0 0 0	0 0 0	0 0 0 0	.00 .00 .00 .00	.00 .00 .00 .00	0 0 0 0	0 0 0	14 14 14 14
				Intertidal Upstream		6739 1620	1261 1906	161.38 731.77	68.58 254.02	0 0	0 0	0 0	0 0	.00	.00 .00	0 0	0 0	42 14
Princ	e William Sound Su	mmary																
				l Intertidal l Upstream		58962 24512		354.59 643.93	21.69 55.92	1 0	557 203	985 34	382 11	1,56 .14	55.92 64.14	1 0	339 1	1312 421

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Order of Collection	Day of Collection	Treatment ^a	Stream Number	Mean	Std. Error	<u>n</u>
1	1	2	692	0.266	0.0052	24
2	1	1	604	0.257	0.0048	24
3	1	2	628	0.384	0.0041	24
4	1	1	621	0.330	0.0042	24
· 5	2	2	618	0.092	0.0040	24
6	2	1	623	0.118	0.0034	24
7	2	2	682	0.126	0.0035	24
8	2	1	695	0.148	0.0028	24

Appendix B. Estimated mean mortality and corresponding standard errors for pink salmon embryos incubated at the Armin F. Koernig Hatchery in 1995.

^a Treatment; 1 = reference, 2 = oil contaminated